

# **A NOVEL ANDROGEN-INDUCED SUPPRESSOR OF CELL PROLIFERATION AND USES THEREOF**

## **Related Information**

This application claims priority to U.S. provisional Application No. 60/121,461, entitled "A NOVEL ANDROGEN-INDUCED SUPPRESSOR OF CELL PROLIFERATION AND USES THEREOF," filed on February 24, 1999, incorporated herein in its entirety by this reference. The contents of all patents, patent applications, and references cited herein are expressly incorporated by reference in their entireties.

## **Government Sponsored Research**

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## **Background of the Invention**

Among men, carcinoma of the prostate is the second most common cancer and the second most common cause of death from cancer in the United States. Each year over 130,000 men are diagnosed with prostate cancer and over 30,000 will die from the disease ((1992) *MMWR* 41:459). Typically, 61% of all deaths from prostate cancer occur within five years of diagnosis and 88% within ten years (Smart (1997) *Cancer* 80:1835-1844).

Moreover, despite the availability of risk assessment tools, the optimal therapy for treating prostate cancer remains controversial (Small (1998) *Drugs Aging* 13:71-81). For example, although certain markers of prostate cancer progression such as prostate-specific antigen (PSA) have proven valuable in the diagnosis and management of prostate cancer, as currently used, PSA is insufficiently sensitive and specific for early detection or staging of the malignancy (Daher *et al.* (1998) *Clin. Chem. Lab. Med.* 36:671-681).

In addition, in some patients with metastatic disease of the prostate, hormone therapy (e.g., antiandrogen, estrogen, etc.) is frequently used. However, many patients on hormone therapy develop hormone resistance and the management of hormone refractory disease is a major clinical problem (Ismail *et al.* (1997) *Tech. Urol.* 3:16-24). The death of patients from prostate cancer is related to the development of clones of cells capable of multiplying and metastasizing without androgen stimulation. To date, efforts to suppress these cells have been of limited success (Newling (1996) *Eur. Urol. Suppl.* 2:69-74). This is in part

due to the fact that the initial events in the development of prostate cancer are not well understood.

Normally, cell numbers in the prostate gland are regulated by androgens through separate pathways that include a) inhibition of cell death (apoptosis), b) induction of cell proliferation (Step-1), and c) inhibition of cell proliferation (Step-2, proliferation shutoff). In normal tissue, the apoptotic and proliferative activities are minimal and apparently, Step-2 (inhibition of cell proliferation) maintains the integrity of the tissue. Prostate cancer cells evolve when this circuitry fails in the initial or early phases in prostate cancer.

## 10 Summary of the Invention

A hope for managing prostate cancer lies in the earlier detection of the disease using improved diagnostic indicators, and developing markers that will allow for the more efficient and strategic use of hormone therapy, preferably in concert with improvements in the quality of life for patients with prostate cancer.

15 To this end, a novel androgen-induced tumor suppressor gene termed "Androgen Shutoff Gene 3" (AS3) has been discovered. This gene has a role in inhibiting cell proliferation and use as a marker for the efficient diagnosis and treatment of prostate cancer.

The present invention is based, at least in part, on the discovery of a novel androgen-induced tumor suppressor, referred to herein as "Androgen Shutoff Gene 3" or "AS3" nucleic acid and protein molecules. The AS3 molecules of the present invention are useful as targets for developing modulating agents of cell proliferation, particularly cells of the prostate. Accordingly, in one aspect, this invention provides isolated nucleic acid molecules encoding AS3 proteins or biologically active portions thereof, as well as nucleic acid fragments suitable as primers or hybridization probes for the detection of AS3-encoding nucleic acids.

In one embodiment, an AS3 nucleic acid molecule of the invention is at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or more identical to the nucleotide sequence (e.g., to the entire length of the nucleotide sequence) shown in SEQ ID NO:1 or 3 or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a complement thereof.

In a preferred embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:1 or 3, or a complement thereof. In another

embodiment, the nucleic acid molecule includes SEQ ID NO:3 and nucleotides 66-4238 of SEQ ID NO:1.

In another embodiment, an AS3 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2 or an amino acid sequence encoded by the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. In a preferred embodiment, an AS3 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to the entire length of the amino acid sequence of SEQ ID NO:2 or the amino acid sequence encoded by the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. In a related embodiment, the nucleic acid encodes a polypeptide fragment having at least 1391 amino acid residues of the amino acid sequence shown in SEQ ID NO: 2.

In another preferred embodiment, an isolated nucleic acid molecule encodes the amino acid sequence of human AS3. In yet another preferred embodiment, the nucleic acid molecule includes a nucleotide sequence encoding a protein having the amino acid sequence of SEQ ID NO:2 or the amino acid sequence encoded by the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_.

Another embodiment of the invention features nucleic acid molecules, preferably AS3 nucleic acid molecules, which specifically detect AS3 nucleic acid molecules relative to nucleic acid molecules encoding non-AS3 proteins. For example, in one embodiment, such a nucleic acid molecule is at least 100-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, or 550-600 or more nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising the nucleotide sequence shown in SEQ ID NO:1, the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a complement thereof. In a related embodiment, the nucleic acid molecule can further contain a nucleotide sequence encoding a heterologous polypeptide.

In a related embodiment, the nucleic acid molecules are at least 15 (e.g., contiguous) nucleotides in length and hybridize under stringent conditions to nucleotides 1-5253 of SEQ ID NO:1.

In other preferred embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2 or an amino acid sequence encoded by the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, wherein the nucleic acid molecule hybridizes to a  
5 nucleic acid molecule comprising SEQ ID NO:1 or 3 under stringent conditions.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to an AS3 nucleic acid molecule, e.g., the coding strand of an AS3 nucleic acid molecule.

Another aspect of the invention provides a vector comprising an AS3 nucleic acid  
10 molecule. In certain embodiments, the vector is a recombinant expression vector. In another embodiment, the invention provides a host cell containing a vector of the invention. In yet another embodiment, the invention provides a host cell containing a nucleic acid molecule of the invention. The invention also provides a method for producing a protein, preferably an AS3 protein, by culturing in a suitable medium, a host cell, e.g., a mammalian  
15 host cell such as a non-human mammalian cell, of the invention containing a recombinant expression vector, such that the protein is produced.

Another aspect of this invention features isolated or recombinant AS3 proteins and polypeptides. In one embodiment, the isolated protein, preferably an AS3 protein has an amino acid sequence at least about 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%,  
20 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2 or the amino acid sequence encoded by the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_.

In another embodiment, the invention features fragments of the AS3 protein having the amino acid sequence of SEQ ID NO:2, wherein the fragment comprises at least 15  
25 amino acids (e.g., contiguous amino acids) of the amino acid sequence of SEQ ID NO:2 or an amino acid sequence encoded by the DNA insert of the plasmid deposited with the ATCC as Accession Number \_\_\_\_\_. In another embodiment, the protein, preferably an AS3 protein, has the amino acid sequence of SEQ ID NO:2, respectively.

In another embodiment, the invention features an isolated protein, preferably an AS3  
30 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence at least about 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to a nucleotide sequence of SEQ ID NO:1 or 3, or a complement thereof. This

invention further features an isolated protein, preferably an AS3 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or 3, or a complement thereof.

5           The proteins of the present invention or portions thereof, e.g., biologically active portions thereof, can be operatively linked to a non-AS3 polypeptide (e.g., heterologous amino acid sequences) to form fusion proteins. The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind proteins of the invention, preferably AS3 proteins.

10           In another aspect, the present invention provides a method for detecting the presence of an AS3 polypeptide in a biological sample by contacting the biological sample with a compound capable of detecting an AS3 polypeptide. In one embodiment of this invention, the compound is an antibody. In another embodiment of this invention, a kit is featured that contains a compound that selectively binds to the polypeptide and instructions for use.

15           In another aspect, the present invention features a method for detecting the presence of an AS3 nucleic acid in a biological sample by contacting the biological sample with a nucleic acid probe or primer that selectively hybridizes to an AS3 molecule of the invention and indicating the presence of such a molecule. In other embodiments of this invention, the nucleic acid in the biological sample is RNA. In another embodiment of the invention, a kit  
20 is provided comprising at least one reagent that binds a nucleic acid and instructions for use.

In another aspect, the invention provides for a method for identifying a compound that binds to an AS3 polypeptide or fragment by detecting direct binding of the compound, binding using a competition assay, or binding using an AS3 activity.

In yet another aspect, the invention provides a method for identifying a compound  
25 that modulates the activity of an AS3 polypeptide or fragment by contacting such a polypeptide or cell expressing such a polypeptide and determining the effect of the test compound on the activity of the polypeptide.

In another aspect, the invention provides a method for modulating AS3 activity comprising contacting a cell capable of expressing AS3 with a compound that modulates  
30 AS3 activity such that AS3 activity in the cell is modulated. In one embodiment, the agent inhibits AS3 activity. In another embodiment, the agent stimulates AS3 activity. In one

embodiment, the compound modulates expression of AS3 by modulating transcription of an AS3 gene or translation of an AS3 mRNA.

In another aspect, the invention features a transgenic animal generated from a cell genetically engineered to lack nucleic acid encoding an AS3 polypeptide, where the  
5 transgenic animal lacks expression of the AS3 polypeptide.

In a related aspect, the invention features a transgenic animal generated from a cell that contains a substantially pure nucleic acid encoding a AS3 polypeptide, where the nucleic acid is expressed in the transgenic animal.

In another aspect, the invention features a method of identifying a compound that  
10 modulates cell proliferation. The method includes: (a) providing a cell that has an AS3 gene; (b) contacting the cell with a candidate compound; and (c) monitoring expression of the AS3 gene, where an alteration in the level of expression of the AS3 gene indicates the presence of a compound which modulates cell proliferation. In one preferred embodiment of this aspect, the alteration that is an increase indicates the compound is inhibiting cell  
15 proliferation, and the alteration that is a decrease indicates the compound is increasing cell proliferation

In a related aspect, the invention features another method of identifying a compound that is able to modulate cell proliferation that includes: (a) providing a cell including a reporter gene operably linked to a promoter from an AS3 gene; (b) contacting the cell with a  
20 candidate compound; and (c) measuring expression of the reporter gene, where a change in the expression in response to the candidate compound identifies a compound that is able to modulate cell proliferation. In one preferred embodiment of this aspect, the alteration that is an increase indicates the compound is inhibiting cell proliferation.

In another aspect, the invention features a method of inhibiting the proliferation of a  
25 cell by administering an amount of AS3 polypeptide or fragment thereof sufficient to inhibit cell proliferation.

In related aspects, the invention includes methods of decreasing cell proliferation by either providing a transgene encoding a AS3 polypeptide or fragment thereof to a cell of an animal such that the transgene is positioned for expression in the cell; or by administering to  
30 the cell a compound which increases AS3 biological activity in a cell. In preferred embodiments, AS3 is from a mammal, the cell being treated is in a mammal, and the

mammal has been diagnosed with a condition involving cell proliferation such as cancer (e.g., prostate cancer).

In two other aspects, the invention features methods of diagnosing a mammal for the presence of disease involving altered cell proliferation or an increased likelihood of developing a disease involving altered cell proliferation. The methods include isolating a sample of nucleic acid from the mammal and determining whether the nucleic acid includes a AS3 mutation, where the presence of a mutation is an indication that the animal has a cell proliferation disease or an increased likelihood of developing a disease involving cell proliferation; or measuring AS3 gene expression in a sample from an animal to be diagnosed, where an alteration in the expression or activity relative to a sample from an unaffected mammal is an indication that the mammal has a disease involving cell proliferation or increased likelihood of developing such a disease. In preferred embodiments, AS3 gene expression is measured by assaying the amount of AS3 polypeptide or AS3 biological activity in the sample (e.g., the AS3 polypeptide is measured by immunological methods), or AS3 gene expression is measured by assaying the amount of AS3 RNA in the sample. In other preferred embodiments, the mammal is a human and the method may be performed after or during hormone therapy (e.g., androgen therapy).

In another aspect, the invention features a kit for diagnosing a mammal for the presence of a disease involving altered cell proliferation or an increased likelihood of developing a disease involving altered cell proliferation that includes a substantially pure antibody that specifically binds a AS3 polypeptide. Another such kit includes material for measuring AS3 RNA (e.g., a probe). In a preferred embodiment, the material is a nucleic acid probe.

In a yet another aspect, the invention features a method of obtaining a AS3 polypeptide, including: (a) providing a cell with DNA encoding a AS3 polypeptide, the DNA being positioned for expression in the cell; (b) culturing the cell under conditions for expressing the DNA; and (c) isolating the AS3 polypeptide.

In a related aspect, the invention features a method of isolating an AS3 gene or portion thereof having sequence identity to human AS3. The method includes amplifying by polymerase chain reaction the AS3 gene or portion thereof using oligonucleotide primers wherein the primers (a) are each greater than 15 nucleotides in length; (b) each have regions of complementarity to opposite DNA strands in a region of the nucleotide sequence

provided in SEQ ID NO: 1; and (c) optionally contain sequences capable of producing restriction endonuclease cut sites in the amplified product; and isolating the AS3 gene or portion thereof.

In another aspect of the invention, the invention features a method for detecting if a subject is at increased risk for developing prostate cancer including the steps of (a) detecting the presence of an AS3 nucleic acid or polypeptide and (b) observing whether or not a subject has reduced or absent AS3 levels as compared to a standard, e.g., normal age matched control, wherein said reduced or absent AS3 levels indicate that the subject is at an increased risk for developing prostate cancer. In a related embodiment, the invention features a kit that contains at least one reagent for detecting the presence of an AS3 molecule.

In another aspect, the invention provides a method of prognosis for prostate cancer by obtaining a biological sample from the subject, measuring AS3 levels, correlating those levels with a control, and determining a prognosis based on whether the subject's AS3 levels are above average or below average. In related embodiments, the method may be performed during or after hormone therapy (e.g., androgen therapy) and employs an antibody or nucleic acid probe to an AS3 molecule.

In even another aspect, the invention features a method for the treatment of prostate cancer comprising identifying a subject with prostate cancer or about to have prostate cancer, administering a hormone therapy, and determining if the subject exhibits a change in AS3 levels. In preferred embodiments, the invention provides a method for identifying subjects that exhibit increased AS3 levels after receiving hormone as responsive to hormone therapy and further, as candidates for intermittent hormone therapy. In other preferred embodiments, the hormone therapy is an androgen therapy, the subject is a human, and the method for measuring includes measuring AS3 nucleic acid or polypeptide levels is performed with an antibody or nucleic acid probe or primer.

Other features and advantages of the invention will be apparent from the following detailed description and claims.



### **Brief Description of the Drawings**

**Figure 1** depicts the cDNA sequence and predicted amino acid sequence of human AS3. The nucleotide sequence corresponds to 1 to 5253 of SEQ ID NO: 1. The amino acid sequence corresponds to amino acids 1 to 1391 of SEQ ID NO: 2. The coding region without the 5' and 3' untranslated regions of the human AS3 gene is shown in SEQ ID NO: 3. Numbers on the left indicate positions in base pairs. The amino acid sequence of the open reading frame is depicted under the coding strand. Numbers on the right indicate amino acid positions. Destabilizing signals found in the untranslated regions of AS3 are underlined and the polyadenylation signal and cleavage site are at base pair positions 5228-5233 and 5249-5253, respectively.

**Figure 2** depicts the N-terminal leucine repeat structure of the AS3 polypeptide. Numbers above the AS3 sequence indicate the positions of the blocks where uninterrupted leucine (or isoleucine, valine) heptades occur.

**Figure 3** depicts sequence comparisons of the putative Mg-nucleotide triphosphate binding subdomains of AS3 with corresponding subdomains of various protein kinases. The boxes represent the Hanks' conserved subdomains, as indicated above each box. The top lines within the boxes show the consensus  $\beta$ -strand, loop, and  $\alpha$ -helical secondary structure elements. The numbers in the second lines indicate the positions of the corresponding conformations in the AS3 sequence. The actual AS3 motifs are shown in the third line. Hanks' conserved subdomains from protein kinases of close similarity are represented in the lines below the AS3 sequence. The names of the kinases are indicated in parentheses. Identical residues are highlighted. In the Mg-ATP binding loop, the "x" and lower case letters indicate non-conserved amino acids.

**Figure 4** depicts the genomic, cosmid, and exon maps of the AS3 cDNA. The chromosomal panel represents a 1 megabase (Mb) genomic region around BRCA2. Boxes with CG numbers are genomic areas where expression of transcripts were detected. The centromer is at the left. The P1 artificial chromosome (PAC) PAC26H23 (Accession No.: Z84467) overlaps with cosmid 267p19 (Accession No.: Z75889), which, in turn, overlaps with PAC49J10 (Accession No.: Z84572). Numbers below the PAC and cosmid lines indicate positions within the genomic clone. The scale above the exon map indicates the genomic distance in thousands of base pairs. In the exon panel, black boxes represent the

exons, while the numbers above them indicate exon numbers. In the mRNA panel, the numbers indicate nucleotide positions.

**Figure 5** depicts the genomic and cDNA positions of exons in the AS3 transcript. Asterisks represent the exon-intron boundaries. The area between asterisks represents the exons. Exon sequences are in upper case, the numbers represent cDNA positions. Lower case letters are intron sequences. Numbers of the first exon indicate positions in PAC26H23. Numbers in parenthesis refer to positions on cosmid 267p19, while numbers in brackets refer to PAC49J10 positions.

**Figure 6.** depicts the cDNA sequence and predicted amino acid sequence of human AS3 having an additional 84 base pairs of untranslated 5' sequence as compared to the sequence presented in Fig. 1. The nucleotide sequence corresponds to 1 to 5337 of SEQ ID NO: 4. The amino acid sequence shown corresponds to amino acids 1 to 1391 of SEQ ID NO: 2. The coding region without the 5' and 3' untranslated regions of the human AS3 gene is shown in SEQ ID NO: 3. Numbers on the left indicate positions in base pairs. The amino acid sequence of the open reading frame is depicted under the coding strand. Numbers on the right indicate amino acid positions. Destabilizing signals found in the untranslated regions of AS3 are underlined and the polyadenylation signal and cleavage signal are at base pair positions 5312-5317 and 5333-5337, respectively.

## **Detailed Description of the Invention**

The present invention is based, at least in part, on the discovery of a novel molecule referred to herein as "Androgen Shutoff Gene 3" or "AS3" nucleic acid and protein molecules, which play a role in hormone-induced inhibition of cell proliferation. In one embodiment, the AS3 molecules are capable of modulating cell proliferation, e.g., cancer. In a preferred embodiment, the AS3 molecules are expressed in cells of the prostate and/or function in the cells of the prostate. In another preferred embodiment, the AS3 molecules are expressed in prostate cells when exposed to a hormone, e.g., an androgen, and inhibit cell proliferation.

The AS3 gene was cloned from a subtracted library made from a human prostate carcinoma cell line induced to undergo growth arrest using androgen.

Androgens regulate prostate cell numbers and cell proliferation by three major mechanisms: a) inhibition of cell death (apoptosis), b) induction of cell proliferation (Step-

1), and c) inhibition of cell proliferation (proliferative shutoff, Step-2) (Isaacs (1985) *Prostate* 5:545-557; Bruchovsky *et al.* (1975) *Vit. & Horm.* 33:61-102; Sonnenschein *et al.*, (1989) *Cancer Res.* 49:3474-3481). Androgens affect epithelial and stromal cell types which, in turn, interact in the prostate (Hayward *et al.*, (1997) *Brit. J. Urol. Suppl.* 2: 18-26).

- 5 The human prostate LNCaP-FGC cell line that exhibits hormone responsiveness and is used extensively for endocrine and molecular studies (Sonnenschein *et al.*, (1989) *Cancer Res.* 49:3474-3481; Horoszewicz *et al.*, (1983) *Cancer Res.* 43:1809-1818; Soto *et al.*, (1995) *Oncology Res.* 7: 545-558) was employed herein. Proliferation is inhibited in these cells by sex steroid-stripped (charcoal-dextran treated) human serum (CDHuS) (Sonnenschein *et al.*,  
 10 (1989) *Cancer Res.* 49:3474-3481). Low androgen concentrations cancel this inhibition (Step-1) and at higher levels, androgens induce an irreversible proliferative shutoff (Step-2) (Sonnenschein *et al.*, (1989) *Cancer Res.* 49:3474-3481; Soto *et al.*, (1995) *Oncology Res.* 7: 545-558). During the shutoff period, these cells remain in the G<sub>0</sub>/G<sub>1</sub> phase of the cell cycle. Prostate specific antigen (PSA) induction, however, is still dependent on androgens  
 15 in these postmitotic cells (Soto *et al.*, (1995) *Oncology Res.* 7: 545-558).

- To dissect androgen-mediated cell-cycle events, several androgen target cell lines that express only one of the steps of the androgen regulated proliferative response were generated. Two LNCaP variants were isolated: the LNCaP-TAC variant which expresses Step-1 only, and the LNCaP-TJA variant, which is resistant to the inhibitory effect of both  
 20 CD serum and androgens (Soto *et al.*, (1995) *Oncology Res.* 7: 545-558). LNCaP-LNO cells, established by Horoszewicz *et al.*, proliferate maximally in the presence of CDHuS, express an androgen-induced proliferative shutoff, and undergo G<sub>0</sub>/G<sub>1</sub> arrest (Step-2) at high androgen concentrations (Horoszewicz *et al.*, (1983) *Cancer Res.* 43:1809-1818; Soto *et al.*, (1995) *Oncology Res.* 7: 545-558). In addition to these human prostate cells, a new model  
 25 to demonstrate the shutoff effect in another cell type was developed by stable transfection of a wild type androgen-receptor construct into breast carcinoma MCF-7 cells. These MCF7-AR1 cells are also able to evoke a proliferative shutoff in response to androgens (Szelei *et al.*, (1997) *Endocrinology* 138: 1406-1412).

- Using a differential subtractive amplification procedure, a set of genes induced in the  
 30 proliferative shutoff response (Step-2) (Wang *et al.*, (1991) *Proc. Natl. Acad. Sci. USA* 88: 11505-11509) was identified. In particular, a gene involved in this regulation, AS3 (androgen shutoff gene 3), that shows high expression in the early regulatory phase of

androgen-induced proliferative shutoff in cell culture and in the prostates of castrated rats was identified.

The AS3 gene encodes a polypeptide of 1391-residues and has a molecular weight of about 186 kD. It has coiled-coil structures that usually participate in protein-protein interactions, a perfect leucine-zipper that suggests DNA binding, nuclear localization motifs, proline- and serine-rich domains, unique C-terminal acidic-basic repeats, and ATP- and DNA-binding motifs.

The transcript has 34 exons in a 200,000 bp region on chromosome 13q12-q13, downstream of the breast cancer susceptibility gene BRCA2, and centromeric to the retinoblastoma (Rb1) locus. This area is subject to frequent allelic losses in cancers, and is believed to carry a number of cryptic suppressor genes.

The AS3 gene is involved in the regulation of androgen-induced proliferative arrest in human prostate cells. Accordingly, the AS3 molecules described below serve as useful diagnostic markers or therapeutic agents to control conditions of aberrant cell proliferation, such as cancer (e.g., cancer of the prostate).

In one embodiment, the present invention is directed at human AS3, however, AS3 family members are also intended to be within the scope of the invention. The term "family" when referring to the protein and nucleic acid molecules of the invention is intended to mean two or more proteins or nucleic acid molecules having a common structural domain or motif and having sufficient amino acid or nucleotide sequence homology as defined herein. Such family members can be naturally or non-naturally occurring and can be from either the same or different species. For example, a family can contain a first protein of human origin, as well as other, distinct proteins of human origin or alternatively, can contain homologues of non-human origin. Members of a family may also have common functional characteristics.

To identify the presence of important domains in a given polypeptide, and make the determination that a protein of interest has a particular profile, the amino acid sequence of the protein can be searched against several databases as described in Example 3. Using these tools, a number of important domains within the AS3 polypeptide have been identified and these results are set forth in Figures 2 and 3 and further described in Example 3.

Isolated proteins of the present invention, preferably AS3 proteins, have an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2 or are encoded

by a nucleotide sequence sufficiently homologous to SEQ ID NO:1 or 3. As used herein, the term "sufficiently homologous" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain) amino acid residues or nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences share common structural domains or motifs and/or a common functional activity. For example, amino acid or nucleotide sequences which share common structural domains have at least 30%, 40%, or 50% homology, preferably 60% homology, more preferably 70%-80%, and even more preferably 90-95% homology across the amino acid sequences of the domains and contain at least one and preferably two structural domains or motifs, are defined herein as sufficiently homologous. Furthermore, amino acid or nucleotide sequences which share at least 30%, 40%, or 50%, preferably 60%, more preferably 70-80%, or 90-95% homology and share a common functional activity are defined herein as sufficiently homologous.

As used interchangeably herein, an "AS3 activity", "biological activity of AS3" or "functional activity of AS3", refers to an activity exerted by an AS3 protein, polypeptide or nucleic acid molecule on an AS3 responsive cell or on an AS3 protein substrate, as determined *in vivo*, or *in vitro*, according to standard techniques. In one embodiment, an AS3 activity is a direct activity, such as an association with an AS3-target molecule. As used herein, a "target molecule" or "binding partner" is a molecule with which an AS3 protein binds or interacts in nature, such that AS3-mediated function is achieved. An AS3 target molecule can be a non-AS3 molecule or an AS3 protein or polypeptide of the present invention. Alternatively, an AS3 activity is an indirect activity, such as modulating cell cycle events. Preferably, an AS3 activity is the ability to modulate androgen-mediated cell proliferation.

Accordingly, another embodiment of the invention features isolated AS3 proteins and polypeptides having an AS3 activity. Preferred proteins are AS3 proteins encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1 or 3.

The nucleotide sequence of the isolated human AS3 cDNA and the predicted amino acid sequence of the human AS3 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively. A plasmid containing the nucleotide sequence encoding human AS3

was deposited with the American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, on \_\_\_\_\_ and assigned Accession Number \_\_\_\_\_. This deposit will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. This  
5 deposit was made merely as a convenience for those of skill in the art and is not an admission that a deposit is required under 35 U.S.C. §112.

The human AS3 gene, which is approximately 5253 nucleotides in length, encodes a protein having a molecular weight of approximately 186 kD and which is approximately 1391 amino acid residues in length.

10 Various aspects of the invention are described in further detail in the following subsections:

#### I. Isolated Nucleic Acid Molecules

One aspect of the invention pertains to isolated nucleic acid molecules that encode  
15 AS3 proteins or biologically active portions thereof, as well as nucleic acid fragments sufficient for use as hybridization probes to identify AS3-encoding nucleic acid molecules (e.g., AS3 mRNA) and fragments for use as PCR primers for the amplification or mutation of AS3 nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (e.g., cDNA or genomic DNA) and RNA molecules (e.g.,  
20 mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

The term "isolated nucleic acid molecule" includes nucleic acid molecules which are separated from other nucleic acid molecules which are present in the natural source of the  
25 nucleic acid. For example, with regards to genomic DNA, the term "isolated" includes nucleic acid molecules which are separated from the chromosome with which the genomic DNA is naturally associated. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived.  
30 For example, in various embodiments, the isolated AS3 nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic

acid is derived (see Fig. 4). Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

5           A nucleic acid molecule of the present invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a portion thereof, can be isolated using standard molecular biology techniques and the sequence information provided herein. Using all or portion of the nucleic acid sequence of SEQ ID  
10 NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, as a hybridization probe, AS3 nucleic acid molecules can be isolated using standard hybridization and cloning techniques (e.g., as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual*.  
15 *2nd, ed., Cold Spring Harbor Laboratory*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989). In a related embodiment, the invention features an AS3 nucleic acid molecule having the sequence of SEQ ID NO: 4 (see Fig. 6) which is identical to the AS3 sequence provided in SEQ ID NO:1 (see Fig. 1) except for an additional 84 base pairs at the 5' end of the molecule. One skilled in the art would recognize that this additional untranslated 5' sequence may indicate that an alternative start site for the beginning of  
20 transcription of the AS3 mRNA molecule may exist.

Moreover, a nucleic acid molecule encompassing all or a portion of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_ can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence of SEQ ID NO:1 or 3,  
25 or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be  
30 cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to AS3 nucleotide sequences can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:1. The sequence of SEQ ID NO:1 corresponds to the human AS3 cDNA. This cDNA comprises sequences encoding the human AS3 protein (i.e., "the coding region", from nucleotides 66-4238), as well as 5' untranslated sequences (nucleotides 1-65) and 3' untranslated sequences (nucleotides 4239-5253). Alternatively, the nucleic acid molecule can comprise only the coding region of SEQ ID NO:1 (e.g., nucleotides 66-4238, corresponding to SEQ ID NO:3).

In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a portion of any of these nucleotide sequences. A nucleic acid molecule which is complementary to the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, is one which is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, such that it can hybridize to the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, thereby forming a stable duplex.

In still another preferred embodiment, an isolated nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least about 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to the entire length of the nucleotide sequence shown in SEQ ID NO:1 or 3, or the entire length of the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a portion of any of these nucleotide sequences.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, for example, a fragment which can be used as a probe or primer or a fragment encoding a portion of an AS3 protein, e.g., a biologically active portion of an AS3 protein. The nucleotide sequence determined from the cloning of the AS3 gene allows for the generation of probes and primers designed for use in identifying and/or cloning other AS3 family members, as well as



AS3 homologues from other species. The probe/primer typically comprises substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12 or 15, preferably about 20 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, or 75 consecutive

- 5 nucleotides of a sense sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, of an anti-sense sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or of a naturally occurring allelic variant or mutant of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert
- 10 of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. In an exemplary embodiment, a nucleic acid molecule of the present invention comprises a nucleotide sequence which is greater than 100-150, 150-200, 200-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, 550-600, 600-650, 650-700, 700-750, 750-800, 800-850, 850-900, 900-950, 950-1000, or more nucleotides in length and hybridizes under stringent
- 15 hybridization conditions to a nucleic acid molecule of SEQ ID NO: 1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_.

Probes based on the AS3 nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments,

20 the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress an AS3 protein, such as by measuring a level of an AS3-encoding nucleic acid in a sample of cells from a subject e.g., detecting AS3 mRNA levels or determining whether a genomic

- 25 AS3 gene has been mutated or deleted.

A nucleic acid fragment encoding a "biologically active portion of an AS3 protein" can be prepared by isolating a portion of the nucleotide sequence of SEQ ID NO: 1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, which encodes a polypeptide having an AS3 biological activity

30 (the biological activities of the AS3 proteins are described herein), expressing the encoded portion of the AS3 protein (e.g., by recombinant expression *in vitro*) and assessing the activity of the encoded portion of the AS3 protein.

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, due to degeneracy of the genetic code and thus encode the same AS3 proteins as those encoded by the nucleotide sequence shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2.

In addition to the AS3 nucleotide sequences shown in SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the AS3 proteins may exist within a population (e.g., the human population). Such genetic polymorphism in the AS3 genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which include an open reading frame encoding an AS3 protein, preferably a mammalian AS3 protein, and can further include non-coding regulatory sequences, and introns.

Allelic variants of human AS3 include both functional and non-functional AS3 proteins. Functional allelic variants are naturally occurring amino acid sequence variants of the human AS3 protein that maintain the ability to modulate cell proliferation, e.g., androgen-induced changes in cell proliferation. Functional allelic variants will typically contain only conservative substitution of one or more amino acids of SEQ ID NO:2 or substitution, deletion or insertion of non-critical residues in non-critical regions of the protein.

Non-functional allelic variants are naturally occurring amino acid sequence variants of the human AS3 protein that do not have the ability to modulate cell proliferation, for example, hormone-induced changes in cell proliferation. Non-functional allelic variants will typically contain a non-conservative substitution, a deletion, or insertion or premature truncation of the amino acid sequence of SEQ ID NO:2 or a substitution, insertion or deletion in critical residues or critical regions.

The present invention further provides non-human orthologues of the human AS3 protein. Orthologues of the human AS3 protein are proteins that are isolated from non-

human organisms and possess the same AS3 ability to modulate cell proliferation, for example, hormone-induced changes in cell proliferation. Orthologues of the human AS3 protein can readily be identified as comprising an amino acid sequence that is substantially homologous to SEQ ID NO:2.

5           Moreover, nucleic acid molecules encoding other AS3 family members and, thus, which have a nucleotide sequence which differs from the AS3 sequences of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_ are intended to be within the scope of the invention. For example, another AS3 cDNA can be identified based on the nucleotide sequence of human AS3.

10          Moreover, nucleic acid molecules encoding AS3 proteins from different species, and which, thus, have a nucleotide sequence which differs from the AS3 sequences of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_ are intended to be within the scope of the invention. For example, a mouse AS3 cDNA can be identified based on the nucleotide sequence of a human AS3.

15           Nucleic acid molecules corresponding to natural allelic variants and homologues of the AS3 cDNAs of the invention can be isolated based on their homology to the AS3 nucleic acids disclosed herein using the cDNAs disclosed herein, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. Nucleic acid molecules corresponding to natural allelic variants  
20          and homologues of the AS3 cDNAs of the invention can further be isolated by mapping to the same chromosome or locus as the AS3 gene.

            Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15, 20, 25, 30 or more nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ  
25          ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. In other embodiment, the nucleic acid is at least 30, 50, 100, 150, 200, 250, 253, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, or 950 nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which  
30          nucleotide sequences at least 60% homologous to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 70%, more preferably at least about 80%, even more preferably at least about 85% or 90% homologous

to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50°C, preferably at 55°C, more preferably at 60°C, and even more preferably at 65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO:1 or 3 corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In addition to naturally-occurring allelic variants of the AS3 sequences that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequences of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, thereby leading to changes in the amino acid sequence of the encoded AS3 proteins, without altering the functional ability of the AS3 proteins. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of AS3 (e.g., the sequence of SEQ ID NO:2) without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the AS3 proteins of the present invention, e.g., those present in the heptad repeat or kinase domains, are predicted to be particularly recalcitrant to alteration. Furthermore, additional amino acid residues that are conserved between the AS3 proteins of the present invention and other members of the ASIC family are not likely to be amenable to alteration.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding AS3 proteins that contain changes in amino acid residues that are not essential for activity. Such AS3 proteins differ in amino acid sequence from SEQ ID NO:2, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid

sequence at least about 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to SEQ ID NO:2.

An isolated nucleic acid molecule encoding an AS3 protein homologous to the protein of SEQ ID NO:2 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_ by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in an AS3 protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of an AS3 coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for AS3 biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO: 1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, the encoded protein can be expressed recombinantly and the activity of the protein can be determined.

In a preferred embodiment, a mutant AS3 protein can be assayed for the ability to ability to modulate cell proliferation, for example, hormone-induced changes in cell proliferation.

In addition to the nucleic acid molecules encoding AS3 proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire AS3 coding strand, or to only a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding AS3. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (e.g., the coding region of human AS3 corresponds to SEQ ID NO:3). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding AS3. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding AS3 disclosed herein (e.g., SEQ ID NO:3), antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of AS3 mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding or noncoding region of AS3 mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of AS3 mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-

2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (i.e., RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an AS3 protein to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention include direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an  $\alpha$ -anomeric nucleic acid molecule. An  $\alpha$ -anomeric nucleic acid molecule forms specific

double-stranded hybrids with complementary RNA in which, contrary to the usual  $\beta$ -units, the strands run parallel to each other (Gaultier et al. (1987) *Nucleic Acids. Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave AS3 mRNA transcripts to thereby inhibit translation of AS3 mRNA. A ribozyme having specificity for an AS3-encoding nucleic acid can be designed based upon the nucleotide sequence of an AS3 cDNA disclosed herein (i.e., SEQ ID NO:1 or 3, or the nucleotide sequence of the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_). For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an AS3-encoding mRNA. See, e.g., Cech et al. U.S. Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, AS3 mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, e.g., Bartel, D. and Szostak, J.W. (1993) *Science* 261:1411-1418.

Alternatively, AS3 gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the AS3 (e.g., the AS3 promoter and/or enhancers) to form triple helical structures that prevent transcription of the AS3 gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. et al. (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher, L.J. (1992) *Bioassays* 14(12):807-15.

In yet another embodiment, the AS3 nucleic acid molecules of the present invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acid molecules can be modified to generate peptide nucleic acids (see Hyrup B. et al. (1996) *Bioorganic & Medicinal Chemistry* 4 (1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics,



e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup B. et al. (1996) *supra*; Perry-O'Keefe et al. *Proc. Natl. Acad. Sci.* 93: 14670-675.

PNAs of AS3 nucleic acid molecules can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, for example, inducing transcription or translation arrest or inhibiting replication. PNAs of AS3 nucleic acid molecules can also be used in the analysis of single base pair mutations in a gene, (e.g., by PNA-directed PCR clamping); as 'artificial restriction enzymes' when used in combination with other enzymes, (e.g., S1 nucleases (Hyrup B. (1996) *supra*)); or as probes or primers for DNA sequencing or hybridization (Hyrup B. et al. (1996) *supra*; Perry-O'Keefe *supra*).

In another embodiment, PNAs of AS3 can be modified, (e.g., to enhance their stability or cellular uptake), by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of AS3 nucleic acid molecules can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, (e.g., RNase H and DNA polymerases), to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup B. (1996) *supra*). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup B. (1996) *supra* and Finn P.J. et al. (1996) *Nucleic Acids Res.* 24 (17): 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs, e.g., 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used as a between the PNA and the 5' end of DNA (Mag, M. et al. (1989) *Nucleic Acid Res.* 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn P.J. et al. (1996) *supra*). Alternatively,

chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser, K.H. et al. (1975) *Bioorganic Med. Chem. Lett.* 5: 1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:6553-6556; Lemaitre et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (See, e.g., Krol et al. (1988) *Bio-Techniques* 6:958-976) or intercalating agents. (See, e.g., Zon (1988) *Pharm. Res.* 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, (e.g., a peptide, hybridization triggered cross-linking agent, transport agent, or hybridization-triggered cleavage agent).

## II. Isolated AS3 Proteins

One aspect of the invention pertains to isolated AS3 proteins, and biologically active portions thereof, as well as polypeptide fragments suitable for use as immunogens to raise anti-AS3 antibodies. In one embodiment, native AS3 proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, AS3 proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, an AS3 protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the AS3 protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of AS3 protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of AS3 protein having less than about 30% (by dry weight) of non-AS3 protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-AS3 protein, still more preferably less than about 10% of non-AS3 protein, and most preferably less than about 5% non-AS3 protein. When the AS3 protein or biologically

active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, i.e., culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

- 5           The language "substantially free of chemical precursors or other chemicals" includes preparations of AS3 protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of AS3 protein having less than about 30% (by dry weight) of chemical
- 10 precursors or non-AS3 chemicals, more preferably less than about 20% chemical precursors or non-AS3 chemicals, still more preferably less than about 10% chemical precursors or non-AS3 chemicals, and most preferably less than about 5% chemical precursors or non-AS3 chemicals.

- As used herein, a "biologically active portion" of an AS3 protein includes a fragment
- 15 of an AS3 protein which participates in an interaction between an AS3 molecule and a non-AS3 molecule. Biologically active portions of an AS3 protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the AS3 protein, e.g., the amino acid sequence shown in SEQ ID NO:2, which include less amino acids than the full length AS3 proteins, and exhibit at least one activity of an AS3
- 20 protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the AS3 protein, e.g., the ability to modulate cell proliferation. A biologically active portion of an AS3 protein can be a polypeptide which is, for example, 10, 25, 50, 100, 200 or more amino acids in length. Biologically active portions of an AS3 protein can be used as targets for developing agents which modulate an AS3 mediated activity, e.g., cell
- 25 proliferation.

- In one embodiment, a biologically active portion of an AS3 protein comprises at least one heptad repeat. It is to be understood that a preferred biologically active portion of an AS3 protein of the present invention may contain two, three, four, or five heptad repeats. As used herein, the term "heptad repeat" includes a protein domain which contains either a
- 30 leucine or similar hydrophobic residue (e.g., isoleucine, valine, or tyrosine) in every seventh position over a stretch of at least 20-30 amino acid residues, more preferably at least 20-25 amino acid residues, and most preferably at least 22 amino acid residues. Typically, the

heptad repeat is uninterrupted, and can participate in protein-protein interactions. The leucine-zipper motif of DNA binding proteins is a specific subclass of this general pattern and is encompassed by the above term. Leucine zipper domains are described in, for example, Landschultz et al. (1988) *Science* 240: 1759-1764, the contents of which are  
 5 incorporated herein by reference. Amino acid residues 55-161, 196-217, 241-277, 319-355, and 375-404 of the AS3 protein all comprise heptad repeats.

In another embodiment, a biologically active portion of an AS3 protein comprises a least one kinase-related domain. It is understood that a preferred biologically active portion of an AS3 protein of the present invention may contain two, three, four, five, six, seven,  
 10 eight, or nine kinase-related domains. As used herein, the term "kinase-related domain" includes a polypeptide consensus motif having high homology to a known protein kinase catalytic domain as described in Hanks *et al.* (1988) *Meth. Enzymol.* 200:38-62 (the contents of which are incorporated herein by reference). In particular, a kinase-related domain is any one of the nine consensus motifs or related sequences set forth in Fig. 3.

15 In another embodiment, a biologically active portion of an AS3 protein can have kinase activity. As referred to herein, "kinase activity" is an activity associated with a protein or polypeptide which is capable of modulating its own phosphorylation state or the phosphorylation state of another protein or polypeptide. Proteins with kinase activity play a role in signaling pathways associated with cellular growth.

20 Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native AS3 protein.

In a preferred embodiment, the AS3 protein has an amino acid sequence shown in SEQ ID NO:2. In other embodiments, the AS3 protein is substantially homologous to SEQ  
 25 ID NO:2, and retains the functional activity of the protein of SEQ ID NO:2, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail in subsection I above. Accordingly, in another embodiment, the AS3 protein is a protein which comprises an amino acid sequence at least about 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98% or more homologous to SEQ ID NO:2.

30 To determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for

optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a preferred embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, or 90% of the length of the reference sequence (e.g., when aligning a second sequence to the AS3 amino acid sequence of SEQ ID NO: 2 having 400 amino acid residues, at least 80, preferably at least 100, more preferably at least 120, even more preferably at least 140, and even more preferably at least 150, 200, 300, or 400 amino acid residues are aligned). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a preferred embodiment, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch (*J. Mol. Biol.* (48):444-453 (1970)) algorithm which has been incorporated into the GAP program in the GCG software package (available at <http://www.gcg.com>), using either a Blossom 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another preferred embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at <http://www.gcg.com>), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. In another embodiment, the percent identity between two amino acid or nucleotide sequences is determined using the algorithm of E. Meyers and W. Miller (*Comput. Appl. Biosci.*, 4:11-17 (1988)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The nucleic acid and protein sequences of the present invention can further be used as a "query sequence" to perform a search against public databases to, for example, identify

other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (1990) *J. Mol. Biol.* 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences homologous to AS3 nucleic acid molecules of the invention. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength = 3 to obtain amino acid sequences homologous to AS3 protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used. See <http://www.ncbi.nlm.nih.gov>.

The invention also provides AS3 chimeric or fusion proteins. As used herein, an AS3 "chimeric protein" or "fusion protein" comprises an AS3 polypeptide operatively linked to a non-AS3 polypeptide. An "AS3 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to AS3, whereas a "non-AS3 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the AS3 protein, e.g., a protein which is different from the AS3 protein and which is derived from the same or a different organism. Within an AS3 fusion protein the AS3 polypeptide can correspond to all or a portion of an AS3 protein. In a preferred embodiment, an AS3 fusion protein comprises at least one biologically active portion of an AS3 protein. In another preferred embodiment, an AS3 fusion protein comprises at least two biologically active portions of an AS3 protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the AS3 polypeptide and the non-AS3 polypeptide are fused in-frame to each other. The non-AS3 polypeptide can be fused to the N-terminus or C-terminus of the AS3 polypeptide.

For example, in one embodiment, the fusion protein is a GST-AS3 fusion protein in which the AS3 sequences are fused to the C-terminus of the GST sequences. Such fusion proteins can facilitate the purification of recombinant AS3.

In another embodiment, the fusion protein is an AS3 protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of AS3 can be increased through use of a heterologous signal sequence.

The AS3 fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject *in vivo*. The AS3 fusion proteins can be used to affect the bioavailability of an AS3 substrate. Use of AS3 fusion proteins may be useful therapeutically for the treatment of disorders caused by, for example, (i) aberrant

- 5 modification or mutation of a gene encoding an AS3 protein; (ii) mis-regulation of the AS3 gene; and (iii) aberrant post-translational modification of an AS3 protein.

Moreover, the AS3-fusion proteins of the invention can be used as immunogens to produce anti-AS3 antibodies in a subject, to purify AS3 ligands and in screening assays to identify molecules which inhibit the interaction of AS3 with an AS3 substrate.

- 10 Preferably, an AS3 chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of
- 15 cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently
- 20 be annealed and reamplified to generate a chimeric gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (e.g., a GST polypeptide). An AS3-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the AS3 protein.

- 25 The present invention also pertains to variants of the AS3 proteins which function as either AS3 agonists (mimetics) or as AS3 antagonists. Variants of the AS3 proteins can be generated by mutagenesis, e.g., discrete point mutation or truncation of an AS3 protein. An agonist of the AS3 proteins can retain substantially the same, or a subset, of the biological activities of the naturally occurring form of an AS3 protein. An antagonist of an AS3
- 30 protein can inhibit one or more of the activities of the naturally occurring form of the AS3 protein by, for example, competitively modulating an AS3-mediated activity of an AS3 protein. Thus, specific biological effects can be elicited by treatment with a variant of

limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the protein has fewer side effects in a subject relative to treatment with the naturally occurring form of the AS3 protein.

5 In one embodiment, variants of an AS3 protein which function as either AS3 agonists (mimetics) or as AS3 antagonists can be identified by screening combinatorial libraries of mutants, e.g., truncation mutants, of an AS3 protein for AS3 protein agonist or antagonist activity. In one embodiment, a variegated library of AS3 variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene  
10 library. A variegated library of AS3 variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential AS3 sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage display) containing the set of AS3 sequences therein. There are a variety of methods which can be used to produce  
15 libraries of potential AS3 variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential AS3 sequences. Methods for synthesizing degenerate  
20 oligonucleotides are known in the art (see, e.g., Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura et al. (1984) *Annu. Rev. Biochem.* 53:323; Itakura et al. (1984) *Science* 198:1056; Ike et al. (1983) *Nucleic Acid Res.* 11:477.

In addition, libraries of fragments of an AS3 protein coding sequence can be used to generate a variegated population of AS3 fragments for screening and subsequent selection of  
25 variants of an AS3 protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of an AS3 coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded  
30 portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be



derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the AS3 protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of AS3 proteins. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recursive ensemble mutagenesis (REM), a new technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify AS3 variants (Arkin and Yourvan (1992) *Proc. Natl. Acad. Sci. USA* 89:7811-7815; Delgrave et al. (1993) *Protein Engineering* 6(3):327-331).

In one embodiment, cell based assays can be exploited to analyze a variegated AS3 library. For example, a library of expression vectors can be transfected into a cell line, e.g., a neuronal cell line, which ordinarily responds to a particular ligand in an AS3-dependent manner. The transfected cells are then contacted with the ligand and the effect of expression of the mutant on signaling by the ligand can be detected, e.g., by measuring intracellular calcium, potassium, or sodium concentration, neuronal membrane depolarization, or the activity of an AS3-regulated transcription factor. Plasmid DNA can then be recovered from the cells which score for inhibition, or alternatively, potentiation of signaling by the ligand, and the individual clones further characterized.

### III Anti-AS3 Antibodies

An isolated AS3 protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind AS3 using standard techniques for polyclonal and monoclonal antibody preparation. A full-length AS3 protein can be used or, alternatively, the invention provides antigenic peptide fragments of AS3 for use as immunogens. The antigenic peptide of AS3 comprises at least 8 amino acid residues of the amino acid sequence shown in SEQ ID NO:2 and encompasses an epitope of AS3 such that

an antibody raised against the peptide forms a specific immune complex with AS3. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues.

- 5 Preferred epitopes encompassed by the antigenic peptide are regions of AS3 that are located on the surface of the protein, e.g., hydrophilic regions, as well as regions with high antigenicity (see, for example, Figure 2).

An AS3 immunogen typically is used to prepare antibodies by immunizing a suitable subject, (e.g., rabbit, goat, mouse or other mammal) with the immunogen. An appropriate  
10 immunogenic preparation can contain, for example, recombinantly expressed AS3 protein or a chemically synthesized AS3 polypeptide. The preparation can further include an adjuvant, such as Freund's complete or incomplete adjuvant, or similar immunostimulatory agent. Immunization of a suitable subject with an immunogenic AS3 preparation induces a polyclonal anti-AS3 antibody response.

- 15 Accordingly, another aspect of the invention pertains to anti-AS3 antibodies. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antigen binding site which specifically binds (immunoreacts with) an antigen, such as AS3. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')<sub>2</sub>  
20 fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind AS3. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of AS3. A monoclonal antibody  
25 composition thus typically displays a single binding affinity for a particular AS3 protein with which it immunoreacts.

Polyclonal anti-AS3 antibodies can be prepared as described above by immunizing a suitable subject with an AS3 immunogen. The anti-AS3 antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked  
30 immunosorbent assay (ELISA) using immobilized AS3. If desired, the antibody molecules directed against AS3 can be isolated from the mammal (e.g., from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG

fraction. At an appropriate time after immunization, e.g., when the anti-AS3 antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) *Nature* 256:495-497) (see also, Brown et al. (1981) *J. Immunol.* 127:539-46; Brown et al. (1980) *J. Biol. Chem.* 255:4980-83; Yeh et al. (1976) *Proc. Natl. Acad. Sci. USA* 76:2927-31; and Yeh et al. (1982) *Int. J. Cancer* 29:269-75), the more recent human B cell hybridoma technique (Kozbor et al. (1983) *Immunol Today* 4:72), the EBV-hybridoma technique (Cole et al. (1985), *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing monoclonal antibody hybridomas is well known (see generally R. H. Kenneth, in *Monoclonal Antibodies: A New Dimension In Biological Analyses*, Plenum Publishing Corp., New York, New York (1980); E. A. Lerner (1981) *Yale J. Biol. Med.*, 54:387-402; M. L. Gefter et al. (1977) *Somatic Cell Genet.* 3:231-36). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with an AS3 immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds AS3.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating an anti-AS3 monoclonal antibody (see, e.g., G. Galfre et al. (1977) *Nature* 266:55052; Gefter et al. *Somatic Cell Genet.*, cited *supra*; Lerner, *Yale J. Biol. Med.*, cited *supra*; Kenneth, *Monoclonal Antibodies*, cited *supra*). Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods which also would be useful. Typically, the immortal cell line (e.g., a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line. Preferred immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a number of myeloma cell lines can be used as a fusion partner according to standard techniques, e.g., the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol

("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for  
5 antibodies that bind AS3, e.g., using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal anti-AS3 antibody can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with AS3 to thereby isolate immunoglobulin library members that bind AS3. Kits for generating and screening phage  
10 display libraries are commercially available (e.g., the Pharmacia *Recombinant Phage Antibody System*, Catalog No. 27-9400-01; and the Stratagene *SurfZAP™ Phage Display Kit*, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, Ladner et al. U.S. Patent No. 5,223,409; Kang et al. PCT International Publication  
15 No. WO 92/18619; Dower et al. PCT International Publication No. WO 91/17271; Winter et al. PCT International Publication WO 92/20791; Markland et al. PCT International Publication No. WO 92/15679; Breitling et al. PCT International Publication WO 93/01288; McCafferty et al. PCT International Publication No. WO 92/01047; Garrard et al. PCT International Publication No. WO 92/09690; Ladner et al. PCT International Publication No.  
20 WO 90/02809; Fuchs et al. (1991) *Bio/Technology* 9:1370-1372; Hay et al. (1992) *Hum. Antibod. Hybridomas* 3:81-85; Huse et al. (1989) *Science* 246:1275-1281; Griffiths et al. (1993) *EMBO J* 12:725-734; Hawkins et al. (1992) *J. Mol. Biol.* 226:889-896; Clarkson et al. (1991) *Nature* 352:624-628; Gram et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:3576-3580; Garrad et al. (1991) *Bio/Technology* 9:1373-1377; Hoogenboom et al. (1991) *Nuc.*  
25 *Acid Res.* 19:4133-4137; Barbas et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:7978-7982; and McCafferty et al. *Nature* (1990) 348:552-554.

Additionally, recombinant anti-AS3 antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention.  
30 Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson et al. International Application No. PCT/US86/02269; Akira, et al. European Patent Application

- 184,187; Taniguchi, M., European Patent Application 171,496; Morrison et al. European Patent Application 173,494; Neuberger et al. PCT International Publication No. WO 86/01533; Cabilly et al. U.S. Patent No. 4,816,567; Cabilly et al. European Patent Application 125,023; Better et al. (1988) *Science* 240:1041-1043; Liu et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:3439-3443; Liu et al. (1987) *J. Immunol.* 139:3521-3526; Sun et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:214-218; Nishimura et al. (1987) *Canc. Res.* 47:999-1005; Wood et al. (1985) *Nature* 314:446-449; and Shaw et al. (1988) *J. Natl. Cancer Inst.* 80:1553-1559; Morrison, S. L. (1985) *Science* 229:1202-1207; Oi et al. (1986) *BioTechniques* 4:214; Winter U.S. Patent 5,225,539; Jones et al. (1986) *Nature* 321:552-525; Verhoeyan et al. (1988) *Science* 239:1534; and Beidler et al. (1988) *J. Immunol.* 141:4053-4060.

An anti-AS3 antibody (e.g., monoclonal antibody) can be used to isolate AS3 by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-AS3 antibody can facilitate the purification of natural AS3 from cells and of recombinantly produced AS3 expressed in host cells. Moreover, an anti-AS3 antibody can be used to detect AS3 protein (e.g., in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the AS3 protein. Anti-AS3 antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, e.g., to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling (i.e., physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include  $^{125}\text{I}$ ,  $^{131}\text{I}$ ,  $^{35}\text{S}$  or  $^3\text{H}$ .

#### IV. Recombinant Expression Vectors

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding an AS3 protein (or a portion thereof). As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (e.g., in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide

sequence in many types of host cells and those which direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, and the like. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., AS3 proteins, mutant forms of AS3 proteins, fusion proteins, and the like).

The recombinant expression vectors of the invention can be designed for expression of AS3 proteins in prokaryotic or eukaryotic cells. For example, AS3 proteins can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith, D.B. and Johnson, K.S. (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Purified fusion proteins can be utilized in AS3 activity assays, (e.g., direct assays or competitive assays described in detail below), or to generate antibodies specific for AS3

proteins, for example. In a preferred embodiment, an AS3 fusion protein expressed in a retroviral expression vector of the present invention can be utilized to infect bone marrow cells which are subsequently transplanted into irradiated recipients. The pathology of the subject recipient is then examined after sufficient time has passed (e.g., six (6) weeks).

5           Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann et al., (1988) *Gene* 69:301-315) and pET 11d (Studier et al., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET  
10   11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to express the  
15   protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada et  
20   al., (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the AS3 expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari, et al., (1987) *Embo J.* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943),  
25   pJRY88 (Schultz et al., (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and picZ (InVitrogen Corp, San Diego, CA).

Alternatively, AS3 proteins can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith et al. (1983) *Mol. Cell Biol.*  
30   3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian



expression vectors include pCDM8 (Seed, B. (1987) *Nature* 329:840) and pMT2PC (Kaufman et al. (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and  
5 Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsch, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.*

In another embodiment, the recombinant mammalian expression vector is capable of  
10 directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the prostate specific promoter (Gotoh *et al.* (1998) *J. Urol.* 60:220-229) albumin promoter (liver-specific; Pinkert et al. (1987) *Genes Dev.* 1:268-277), lymphoid-specific  
15 promoters (Calame and Eaton (1988) *Adv. Immunol.* 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) *EMBO J.* 8:729-733) and immunoglobulins (Banerji et al. (1983) *Cell* 33:729-740; Queen and Baltimore (1983) *Cell* 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) *Proc. Natl. Acad. Sci. USA* 86:5473-5477), pancreas-specific promoters (Edlund et al.  
20 (1985) *Science* 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally-regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) *Science* 249:374-379) and the  $\alpha$ -fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev.* 3:537-546).

25 The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to AS3 mRNA. Regulatory sequences operatively linked to a nucleic acid cloned  
30 in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or

cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a

5 discussion of the regulation of gene expression using antisense genes see Weintraub, H. et al., Antisense RNA as a molecular tool for genetic analysis, *Reviews - Trends in Genetics*, Vol. 1(1) 1986.

#### V. Host Cells

10 Another aspect of the invention pertains to host cells into which an AS3 nucleic acid molecule of the invention is introduced, e.g., an AS3 nucleic acid molecule within a recombinant expression vector or an AS3 nucleic acid molecule containing sequences which allow it to homologously recombine into a specific site of the host cell's genome. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood

15 that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, an AS3 protein

20 can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and

25 "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (*Molecular Cloning: A Laboratory Manual*, 2nd, ed., Cold Spring Harbor

30 *Laboratory*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding an AS3 protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (i.e., express) an AS3 protein. Accordingly, the invention further provides methods for producing an AS3 protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of the invention (into which a recombinant expression vector encoding an AS3 protein has been introduced) in a suitable medium such that an AS3 protein is produced. In another embodiment, the method further comprises isolating an AS3 protein from the medium or the host cell.

## VI. Transgenic Animals

The host cells of the invention can also be used to produce non-human transgenic animals. For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which AS3-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous AS3 sequences have been introduced into their genome or homologous recombinant animals in which endogenous AS3 sequences have been altered. Such animals are useful for studying the function and/or activity of an AS3 and for identifying and/or evaluating modulators of AS3 activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic

animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous AS3 gene  
5 has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing an AS3-encoding nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral  
10 infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The AS3 cDNA sequence of SEQ ID NO:1 can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of a human AS3 gene, such as a mouse or rat AS3 gene, can be used as a transgene. Alternatively, an AS3 gene homologue, such as another AS3 family member, can be isolated based on hybridization to  
15 the AS3 cDNA sequences of SEQ ID NO:1 or 3, or the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_ (described further in subsection I above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to an AS3 transgene to direct expression of an  
20 AS3 protein to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., *Manipulating the Mouse Embryo*, (Cold Spring Harbor Laboratory Press, Cold Spring  
25 Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of an AS3 transgene in its genome and/or expression of AS3 mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding an AS3 protein can  
30 further be bred to other transgenic animals carrying other transgenes.

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of an AS3 gene into which a deletion, addition or substitution has been

introduced to thereby alter, e.g., functionally disrupt, the AS3 gene. The AS3 gene can be a human gene (e.g., the cDNA of SEQ ID NO:3), but more preferably, is a non-human homologue of a human AS3 gene (e.g., a cDNA isolated by stringent hybridization with the nucleotide sequence of SEQ ID NO:1). For example, a mouse AS3 gene can be used to

5 construct a homologous recombination nucleic acid molecule, e.g., a vector, suitable for altering an endogenous AS3 gene in the mouse genome. In a preferred embodiment, the homologous recombination nucleic acid molecule is designed such that, upon homologous recombination, the endogenous AS3 gene is functionally disrupted (i.e., no longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the homologous

10 recombination nucleic acid molecule can be designed such that, upon homologous recombination, the endogenous AS3 gene is mutated or otherwise altered but still encodes functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous AS3 protein). In the homologous recombination nucleic acid molecule, the altered portion of the AS3 gene is flanked at its 5' and 3' ends by additional

15 nucleic acid sequence of the AS3 gene to allow for homologous recombination to occur between the exogenous AS3 gene carried by the homologous recombination nucleic acid molecule and an endogenous AS3 gene in a cell, e.g., an embryonic stem cell. The additional flanking AS3 nucleic acid sequence is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of

20 flanking DNA (both at the 5' and 3' ends) are included in the homologous recombination nucleic acid molecule (see, e.g., Thomas, K.R. and Capecchi, M. R. (1987) *Cell* 51:503 for a description of homologous recombination vectors). The homologous recombination nucleic acid molecule is introduced into a cell, e.g., an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced AS3 gene has homologously recombined

25 with the endogenous AS3 gene are selected (see e.g., Li, E. et al. (1992) *Cell* 69:915). The selected cells can then be injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see e.g., Bradley, A. in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E.J. Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the

30 embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for

constructing homologous recombination nucleic acid molecules, e.g., vectors, or homologous recombinant animals are described further in Bradley, A. (1991) *Current Opinion in Biotechnology* 2:823-829 and in PCT International Publication Nos.: WO 90/11354 by Le Mouellec et al.; WO 91/01140 by Smithies et al.; WO 92/0968 by Zijlstra et al.; and WO 93/04169 by Berns et al.

In another embodiment, transgenic non-humans animals can be produced which contain selected systems which allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman et al. (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, e.g., by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. (1997) *Nature* 385:810-813 and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G<sub>0</sub> phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The reconstructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

## VII. Pharmaceutical Compositions

The AS3 nucleic acid molecules, fragments of AS3 proteins, and anti-AS3 antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a

pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for

5 pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with

10 its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols,

15 glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The

20 parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration,

25 suitable carriers include physiological saline, bacteriostatic water, Cremophor EL™ (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or

30 dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as

lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, 5 for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

10 Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a fragment of an AS3 protein or an anti-AS3 antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for 15 the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral 20 therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The 25 tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring 30 agent such as peppermint, methyl salicylate, or orange flavoring.



Systemic administration can also be by transmucosal or transdermal means. For  
5 transmucosal or transdermal administration, penetrants appropriate to the barrier to be  
permeated are used in the formulation. Such penetrants are generally known in the art, and  
include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid  
derivatives. Transmucosal administration can be accomplished through the use of nasal  
sprays or suppositories. For transdermal administration, the active compounds are  
10 formulated into ointments, salves, gels, or creams as generally known in the art.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular

therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (*i.e.*, the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

### VIII. Gene Therapy

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Patent 5,328,470) or by stereotactic injection (see *e.g.*, Chen et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be

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## IX. Uses and Methods of the Invention

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In one embodiment, the invention provides assays for screening candidate or test compounds which are substrates of an AS3 protein or polypeptide or biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of an AS3 protein or polypeptide or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt *et al.* (1993) *Proc. Natl. Acad. Sci. U.S.A.* 90:6909; Erb *et al.* (1994) *Proc. Natl. Acad. Sci. USA* 91:11422; Zuckermann *et al.* (1994). *J. Med. Chem.* 37:2678; Cho *et al.* (1993) *Science* 261:1303; Carrell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.* 33:2059; Carell *et al.* (1994) *Angew. Chem. Int. Ed. Engl.* 33:2061; and in Gallop *et al.* (1994) *J. Med. Chem.* 37:1233.

Libraries of compounds may be presented in solution (e.g., Houghten (1992) *Biotechniques* 13:412-421), or on beads (Lam (1991) *Nature* 354:82-84), chips (Fodor (1993) *Nature* 364:555-556), bacteria (Ladner USP 5,223,409), spores (Ladner USP '409), plasmids (Cull *et al.* (1992) *Proc Natl Acad Sci USA* 89:1865-1869) or on phage (Scott and Smith (1990) *Science* 249:386-390); (Devlin (1990) *Science* 249:404-406); (Cwirla *et al.* (1990) *Proc. Natl. Acad. Sci.* 87:6378-6382); (Felici (1991) *J. Mol. Biol.* 222:301-310); (Ladner *supra.*).

In one embodiment, an assay is a cell-based assay in which a cell which expresses an AS3 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate AS3 activity is determined. Determining the ability of the test compound to modulate AS3 activity can be determined by monitoring, for example, changes in cell using standard techniques.

In another embodiment, an assay of the present invention is a cell-free assay in which an AS3 protein or biologically active portion thereof is contacted with a test

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compound and the ability of the test compound to bind to the AS3 protein or biologically active portion thereof is determined. Preferred biologically active portions of the AS3 proteins to be used in assays of the present invention include fragments which participate in interactions with non-AS3 molecules, e.g., fragments with high surface probability scores (see, for example, Figure 2). Binding of the test compound to the AS3 protein can be determined either directly or indirectly as described above. In a preferred embodiment, the assay includes contacting the AS3 protein or biologically active portion thereof with a known compound which binds AS3 to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with an AS3 protein, wherein determining the ability of the test compound to interact with an AS3 protein comprises determining the ability of the test compound to preferentially bind to AS3 or biologically active portion thereof as compared to the known compound.

In yet another embodiment, the cell-free assay involves contacting an AS3 protein or biologically active portion thereof with a known compound which binds the AS3 protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the AS3 protein, wherein determining the ability of the test compound to interact with the AS3 protein comprises determining the ability of the AS3 protein to preferentially bind to or modulate the activity of an AS3 target molecule.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize AS3 to facilitate separation of proteins that interact with AS3, as well as to accommodate automation of the assay. Binding of a test compound to an AS3 protein, or interaction of an AS3 protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase/ AS3 fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or AS3 protein, and the mixture incubated under conditions conducive to complex formation (e.g., at physiological

conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above.

- Alternatively, the complexes can be dissociated from the matrix, and the level of AS3  
5 binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either an AS3 protein or an AS3 target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated AS3 protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide)  
10 using techniques known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with AS3 protein or target molecules but which do not interfere with binding of the AS3 protein to its target molecule can be derivatized to the wells of the plate, and unbound target or AS3 protein trapped in the wells by antibody  
15 conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the AS3 protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the AS3 protein or target molecule.

20 In another embodiment, modulators of AS3 expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of AS3 mRNA or protein in the cell is determined. The level of expression of AS3 mRNA or protein in the presence of the candidate compound is compared to the level of expression of AS3 mRNA or protein in the absence of the candidate compound. The candidate compound can then be  
25 identified as a modulator of AS3 expression based on this comparison. For example, when expression of AS3 mRNA or protein is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of AS3 mRNA or protein expression. Alternatively, when expression of AS3 mRNA or protein is less (statistically significantly less) in the presence of  
30 the candidate compound than in its absence, the candidate compound is identified as an inhibitor of AS3 mRNA or protein expression. This assay may be further modified to include the presence of a hormone, e.g., an androgen or an anti-androgen. The level of AS3

mRNA or protein expression in the cells can be determined by methods described herein for detecting AS3 mRNA or protein.

In yet another aspect of the invention, the AS3 proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al. (1993) *Cell* 72:223-232; Madura et al. (1993) *J. Biol. Chem.* 268:12046-12054; Bartel et al. (1993) *Biotechniques* 14:920-924; Iwabuchi et al. (1993) *Oncogene* 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with AS3 ("AS3-binding proteins" or "AS3-bp") and are involved in AS3 activity. Such AS3-binding proteins are also likely to be involved in the propagation of signals by the AS3 proteins or AS3 targets as, for example, downstream elements of an AS3-mediated signaling pathway. Alternatively, such AS3-binding proteins are likely to be AS3 inhibitors.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for an AS3 protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming an AS3-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the AS3 protein.

In another aspect, the invention pertains to a combination of two or more of the assays described herein. For example, a modulating agent can be identified using a cell-based or a cell free assay, and the ability of the agent to modulate the activity of an AS3 protein can be confirmed *in vivo*, e.g., in an animal such as an animal model for pain.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent

identified as described herein (e.g., an AS3 modulating agent, an antisense AS3 nucleic acid molecule, an AS3-specific antibody, or an AS3-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to  
5 determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

#### IX, B, Detection Assays

10 Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue typing); and (iii) aid in forensic  
15 identification of a biological sample. These applications are described in the subsections below.

#### IX, B, 1., Chromosome Mapping

Once the sequence (or a portion of the sequence) of a gene has been isolated, this  
20 sequence can be used to map the location of the gene on a chromosome. This process is called chromosome mapping. Accordingly, portions or fragments of the AS3 nucleotide sequences, described herein, can be used to map the location of the AS3 genes on a chromosome as described in Example 4. The mapping of the AS3 sequences to chromosomes is an important step in correlating these sequences with genes associated with  
25 disease.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the AS3 gene, can be determined. If a mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of  
30 affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete



sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

#### IX, B, 2., Tissue Typing

5           The AS3 sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for identification. This  
10   method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

          Furthermore, the sequences of the present invention can be used to provide an  
15   alternative technique which determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the AS3 nucleotide sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

          Panels of corresponding DNA sequences from individuals, prepared in this manner,  
20   can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The AS3 nucleotide sequences of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a  
25   greater degree in the noncoding regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to  
30   differentiate individuals. The noncoding sequences of SEQ ID NO:1 can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers which each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences,

such as those in SEQ ID NO:3 are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

If a panel of reagents from AS3 nucleotide sequences described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

#### IX, B, 3., Use of Partial AS3 Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means for positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, e.g., hair or skin, or body fluids, e.g., blood, saliva, or semen found at a crime scene. The amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

The sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e. another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Sequences targeted to noncoding regions of SEQ ID NO:1 are particularly appropriate for this use as greater numbers of polymorphisms occur in the noncoding regions, making it easier to differentiate individuals using this technique. Examples of polynucleotide reagents include the AS3 nucleotide sequences or portions thereof, e.g., fragments derived from the noncoding regions of SEQ ID NO:1 having a length of at least 20 bases, preferably at least 30 bases.

The AS3 nucleotide sequences described herein can further be used to provide polynucleotide reagents, e.g., labeled or labelable probes which can be used in, for example, an *in situ* hybridization technique, to identify a specific tissue, e.g., brain tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown

origin. Panels of such AS3 probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, e.g., AS3 primers or probes can be used to screen tissue culture for contamination (i.e. screen for the presence of a mixture of different types  
5 of cells in a culture).

#### IX, C, Predictive Medicine

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic  
10 (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining AS3 protein and/or nucleic acid expression as well as AS3 activity, in the context of a biological sample (e.g., blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant or  
15 reduced AS3 expression or activity. The invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with AS3 protein, nucleic acid expression or activity. For example, mutations in an AS3 gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby prophylactically treat an individual prior to the onset of a  
20 disorder characterized by or associated with AS3 protein, nucleic acid expression, or activity.

Another aspect of the invention pertains to monitoring the influence of agents (e.g., drugs, compounds) on the expression or activity of AS3 in clinical trials.

These and other agents are described in further detail in the following sections and in  
25 Examples 4 and 5.

#### IX, C, 1., Diagnostic Assays

An exemplary method for detecting the presence or absence of AS3 protein or nucleic acid in a biological sample involves obtaining a biological sample from a test subject  
30 and contacting the biological sample with a compound or an agent capable of detecting AS3 protein or nucleic acid (e.g., mRNA, genomic DNA) that encodes AS3 protein such that the presence of AS3 protein or nucleic acid is detected in the biological sample. A preferred

agent for detecting AS3 mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to AS3 mRNA or genomic DNA. The nucleic acid probe can be, for example, a full-length AS3 nucleic acid, such as the nucleic acid of SEQ ID NO:1 or 3, or the DNA insert of the plasmid deposited with ATCC as Accession Number \_\_\_\_\_, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to AS3 mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

A preferred agent for detecting AS3 protein is an antibody capable of binding to AS3 protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')<sub>2</sub>) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect AS3 mRNA, protein, or genomic DNA in a biological sample *in vitro* as well as *in vivo*. For example, *in vitro* techniques for detection of AS3 mRNA include Northern hybridizations and *in situ* hybridizations. *In vitro* techniques for detection of AS3 protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of AS3 genomic DNA include Southern hybridizations.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a tissue or cell sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent

capable of detecting AS3 protein, mRNA, or genomic DNA, such that the presence of AS3 protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of AS3 protein, mRNA or genomic DNA in the control sample with the presence of AS3 protein, mRNA or genomic DNA in the test sample.

- 5           The invention also encompasses kits for detecting the presence of AS3 in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting AS3 protein or mRNA in a biological sample; means for determining the amount of AS3 in the sample; and means for comparing the amount of AS3 in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit  
10 can further comprise instructions for using the kit to detect AS3 protein or nucleic acid.

#### IX, C, 2., Prognostic Assays

- The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant AS3  
15 expression or activity. As used herein, the term "aberrant" includes an AS3 expression or activity which deviates from the wild type AS3 expression or activity. Aberrant expression or activity includes increased or decreased expression or activity, as well as expression or activity which does not follow the wild type developmental pattern of expression or the subcellular pattern of expression. For example, aberrant AS3 expression or activity is  
20 intended to include the cases in which a mutation in the AS3 gene causes the AS3 gene to be under-expressed or over-expressed and situations in which such mutations result in a non-functional AS3 protein or a protein which does not function in a wild-type fashion, e.g., a protein which does not interact with an AS3 ligand or one which interacts with a non-AS3 ligand.

- 25           The assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing a malignancy associated with a misregulation in AS3 protein activity or nucleic acid expression, such as a prostate cancer. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant AS3 expression or activity in which a test sample is  
30 obtained from a subject and AS3 protein or nucleic acid (e.g., mRNA or genomic DNA) is detected, wherein the presence of AS3 protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant or unwanted

AS3 expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest. For example, a test sample can be a cell, tissue, or biological fluid containing a cell.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate, such as a hormone, e.g., an androgen) to treat a disease or disorder associated with aberrant AS3 expression or activity. For example, such methods can be used to determine whether a subject can be effectively treated with an agent for aberrant cell proliferation, e.g., cancer of the prostate.

The methods of the invention can also be used to detect genetic alterations in an AS3 gene, thereby determining if a subject with the altered gene is at risk for a disorder characterized by misregulation in AS3 protein activity or nucleic acid expression, such as a proliferative disorder, e.g., cancer. In preferred embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding an AS3-protein, or the mis-expression of the AS3 gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of 1) a deletion of one or more nucleotides from an AS3 gene; 2) an addition of one or more nucleotides to an AS3 gene; 3) a substitution of one or more nucleotides of an AS3 gene, 4) a chromosomal rearrangement of an AS3 gene; 5) an alteration in the level of a messenger RNA transcript of an AS3 gene, 6) aberrant modification of an AS3 gene, such as of the methylation pattern of the genomic DNA, 7) the presence of a non-wild type splicing pattern of a messenger RNA transcript of an AS3 gene, 8) a non-wild type level of an AS3-protein, 9) allelic loss of an AS3 gene, and 10) inappropriate post-translational modification of an AS3-protein. As described herein, there are a large number of assays known in the art which can be used for detecting alterations in an AS3 gene. A preferred biological sample is a cell, tissue, or biological fluid containing a cell, isolated by conventional means from a subject.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al. (1988) *Science* 241:1077-1080; and Nakazawa et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:360-364), the latter of which can be particularly useful for detecting point

mutations in the AS3-gene (see Abravaya et al. (1995) *Nucleic Acids Res* .23:675-682).

This method can include the steps of collecting a sample of cells from a subject, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the

nucleic acid sample with one or more primers which specifically hybridize to an AS3 gene

5 under conditions such that hybridization and amplification of the AS3-gene (if present)

occurs, and detecting the presence or absence of an amplification product, or detecting the

size of the amplification product and comparing the length to a control sample. It is

anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

10 Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. et al., (1990) *Proc. Natl. Acad. Sci. USA* 87:1874-1878), transcriptional amplification system (Kwoh, D.Y. et al., (1989) *Proc. Natl. Acad. Sci. USA* 86:1173-1177), Q-Beta Replicase (Lizardi, P.M. et al. (1988) *Bio-Technology* 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using

15 techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in an AS3 gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and

20 control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Patent No. 5,498,531) can be used to score for the presence of specific

25 mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in AS3 can be identified by hybridizing a sample and control nucleic acids, e.g., DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin, M.T. et al. (1996) *Human Mutation* 7: 244-255; Kozal, M.J. et al. (1996) *Nature Medicine* 2: 753-759). For example,

30 genetic mutations in AS3 can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. *et al. supra*. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample

and control to identify base changes between the sequences by making linear arrays of sequential overlapping probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the AS3 gene and detect mutations by comparing the sequence of the sample AS3 with the corresponding wild-type (control) sequence.

- Examples of sequencing reactions include those based on techniques developed by Maxam and Gilbert ((1977) *Proc. Natl. Acad. Sci. USA* 74:560) or Sanger ((1977) *Proc. Natl. Acad. Sci. USA* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, e.g., PCT International Publication No. WO 94/16101; Cohen et al. (1996) *Adv. Chromatogr.* 36:127-162; and Griffin et al. (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

- Other methods for detecting mutations in the AS3 gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers et al. (1985) *Science* 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wild-type AS3 sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent which cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton *et al.* (1988) *Proc. Natl Acad Sci USA* 85:4397; Saleeba et al. (1992) *Methods Enzymol.* 217:286-295. In a preferred embodiment, the control DNA or RNA can be labeled for detection.



In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in AS3 cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994) *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on an AS3 sequence, e.g., a wild-type AS3 sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in AS3 genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita *et al.* (1989) *Proc Natl. Acad. Sci USA*: 86:2766, see also Cotton (1993) *Mutat. Res.* 285:125-144; and Hayashi (1992) *Genet. Anal. Tech. Appl.* 9:73-79). Single-stranded DNA fragments of sample and control AS3 nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In a preferred embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen *et al.* (1991) *Trends Genet* 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers *et al.* (1985) *Nature* 313:495). When DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing

gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys Chem* 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki *et al.* (1986) *Nature* 324:163); Saiki *et al.* (1989) *Proc. Natl Acad. Sci USA* 86:6230). Such allele specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology which depends on selective PCR amplification may be used in conjunction with the instant invention. Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs *et al.* (1989) *Nucleic Acids Res.* 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner (1993) *Tibtech* 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini *et al.* (1992) *Mol. Cell Probes* 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) *Proc. Natl. Acad. Sci USA* 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving an AS3 gene.

Furthermore, any cell type or tissue in which AS3 is expressed, *e.g.*, prostate tissue, may be utilized in the prognostic assays described herein.

IX, C, 3., Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (e.g., hormone therapy) on the expression or activity of an AS3 protein (e.g., the modulation of cell proliferation) can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase AS3 gene expression, protein levels, or upregulate AS3 activity, can be monitored in clinical trials of subjects exhibiting decreased AS3 gene expression, protein levels, or downregulated AS3 activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease AS3 gene expression, protein levels, or downregulate AS3 activity, can be monitored in clinical trials of subjects exhibiting increased AS3 gene expression, protein levels, or upregulated AS3 activity. In such clinical trials, the expression or activity of an AS3 gene, and preferably, other genes (e.g., prostate-specific antigen (PSA)) that have been implicated in, for example, an AS3-associated disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes, including AS3, that are modulated in cells by treatment with an agent (e.g., compound, drug or hormone) which modulates AS3 activity (e.g., identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on cell proliferation modulated by AS3, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of AS3 and other genes implicated in the AS3-associated disorder. The levels of gene expression (e.g., a gene expression pattern) can be quantified by northern blot analysis or RT-PCR, as described herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of AS3 or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be determined before, and at various points during treatment of the individual with the agent or may be used to determine when treatment is appropriate.

In a preferred embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate, e.g., a hormone, identified by the screening assays described herein) including the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii)

- detecting the level of expression of an AS3 protein, mRNA, or genomic DNA in the preadministration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the AS3 protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the AS3 protein, mRNA, or genomic DNA in the pre-administration sample with the AS3 protein, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of AS3 to higher levels than detected, i.e., to increase the effectiveness of the agent.
- Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of AS3 to lower levels than detected, i.e. to decrease the effectiveness of the agent. According to such an embodiment, AS3 expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

#### IX, D., Methods of Treatment

The present invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant AS3 expression or activity. With regards to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics. "Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market. More specifically, the term refers to the study of how a patient's genes determine his or her response to a drug (e.g., a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with either the AS3 molecules of the present invention or AS3 modulators according to that individual's drug response genotype. Pharmacogenomics allows a clinician or physician to target prophylactic or therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug-related side effects.

IX, D., 1., Prophylactic Methods

In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with an aberrant AS3 expression or activity, by administering to the subject an AS3 molecule or an agent which modulates AS3 expression or at least AS3 activity. Subjects at risk for a disease (e.g., prostate cancer) which is caused or contributed to by aberrant AS3 expression or activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the AS3 aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of AS3 aberrancy, for example, an AS3 molecule, AS3 agonist (e.g., hormone therapy), or AS3 antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

IX, D., 2., Therapeutic Methods

Another aspect of the invention pertains to methods of modulating AS3 expression or activity for therapeutic purposes. Accordingly, in an exemplary embodiment, the modulatory method of the invention involves contacting a cell with an AS3 molecule or agent that modulates AS3 protein activity associated with cell (e.g., cell proliferation). An agent that modulates AS3 protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of an AS3 protein (e.g., an AS3 substrate), an AS3 antibody, an AS3 agonist or antagonist, a peptidomimetic of an AS3 agonist or antagonist, or other small molecule (e.g., hormone, such as an androgen). In one embodiment, the agent stimulates one or more AS3 activities. Examples of such stimulatory agents include androgen therapy or a nucleic acid molecule encoding AS3 that has been introduced into the cell. These modulatory methods can be performed *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a cell proliferative disease (e.g., prostate cancer) or disorder characterized by aberrant or reduced expression or activity of an AS3 protein or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay or a hormone such as androgen as described herein), or combination of agents that modulates AS3 expression or activity.

Stimulation of AS3 activity is desirable in situations in which AS3 is abnormally downregulated and/or in which increased AS3 activity is likely to have a beneficial effect. For example, stimulation of AS3 activity is desirable in a cell proliferative disease such as prostate cancer and increasing AS3 activity is likely to have a beneficial effect. Moreover, the ability to detect androgen-induced AS3 expression in a patient is an indication that the patient is responsive to hormone and therefore a candidate for intermittent hormone therapy. As used herein, the term "intermittent hormone therapy" or "intermittent hormone treatment" includes a treatment regime wherein a patient is treated with a hormone, such as an androgen, for a period of time and then withdrawn from such treatment for a period of time. This intermittent administration of hormone shall be, at least in part, determined by an analysis of the patients AS3 levels as described herein.

#### IX, D, 3., Pharmacogenomics

The AS3 molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on AS3 activity (e.g., AS3 gene expression) as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) AS3-associated disorders (e.g., cell proliferation) associated with aberrant or reduced AS3 activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer an AS3 molecule or AS3 modulator as well as tailoring the dosage and/or therapeutic regimen of treatment with an AS3 molecule or AS3 modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, for example, Eichelbaum, M. et al. (1996) *Clin. Exp. Pharmacol. Physiol.* 23(10-11):983-985 and Linder, M.W. et al. (1997) *Clin. Chem.* 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic

conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical  
5 complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (e.g., a "bi-allelic" gene  
10 marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants.) Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be  
15 generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a "SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of  
20 such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to  
25 identify genes that predict drug response. According to this method, if a gene that encodes a drugs target is known (e.g., an AS3 protein of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major  
30 determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (e.g., N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why

some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For  
5 example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic  
10 effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to  
15 identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (e.g., an AS3 molecule or AS3 modulator of the present invention) can give an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for  
20 prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with an AS3 molecule or AS3 modulator, such as a modulator identified by one of the exemplary screening assays described herein.

25 This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application are incorporated herein by reference.



## EXAMPLE 1.

ISOLATION AND CLONING OF HUMAN AS3

In this example, the isolation and cloning of the gene encoding human AS3 is described.

5 To isolate a cDNA encoding an inhibitor of prostate cancer progression, the LNCaP-FGC cell line established from a metastatic lymph node from a patient with prostate adenocarcinoma (Horoszewicz *et al.*, (1983) *Cancer Res.* 43:1809-1818) was utilized. This cell line and related cell lines derived therefrom were cultured as previously described (Soto *et al.*, (1995) *Oncology Res.* 7: 545-558; Soto *et al.*, (1991) *J. Steroid Biochem.* 23: 87-94).

10 In addition, to isolate a gene expressed at the proliferative shutoff point (i.e., Step 2), a subtractive strategy was used whereby the proliferation of LNCaP-FGC cells was arrested using two different treatments, namely, CD serum (i.e., androgen free serum) and high androgen concentrations. This selective approach takes advantage of the fact that the cells were equally arrested at the G1 stage of the cell cycle by different mechanisms (Soto *et al.*,  
15 (1995) *Oncology Res.* 7: 545-558).

Moreover, since regulatory mRNAs are frequently expressed at low copy numbers, a protocol was adopted using repeated PCR cycles to selectively amplify these sequences. The final subtracted pool, therefore, was enriched to represent high ranking regulatory elements in the androgen-induced proliferative shutoff (Step-2).

20 | Briefly, androgen-specific, low-abundance regulatory mRNA sequences expressed during the proliferative shutoff, were selected using the Wang-Brown approach (Wang *et al.*, (1991) *Proc. Natl. Acad. Sci. USA* 88: 11505-11509). Short fragments of cDNAs were amplified first: then three cycles of subtractions and amplifications between the control and proliferation arrested cDNAs resulted in sequence pools that were differentially expressed  
25 (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218). LNCaP-FGC cells were treated with 30 nM R1881 to generate proliferative shutoff. R1881 (methyltrienolone) is a synthetic, non-metabolized androgen (Roussel-UCLAF, Romainville, France). Exposure to androgen for 24 hours was required to commit LNCaP-FGC cells to an irreversible proliferative shutoff (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218). It was  
30 concluded that at this point, the genes responsible for the shutoff were highly induced. LNCaP-FGC cells reversibly arrested by CDHuS were considered as the shutoff-negative control; they were harvested after three days of CDHuS treatment. Total RNA was prepared

AS3  
A3

by the acidic guanidinium-thiocyanate method and polyA<sup>+</sup> RNA was purified by using the FastTrack kit (Invitrogen, San Diego, CA) (Chomczynsky *et al.*, (1987) *Anal. Biochem.* 162: 156-159).

Double-stranded cDNA pools from R1881-treated cells (R cDNA) and CDHuS- treated cells (CD cDNA) were synthesized using the Copy Kit (Invitrogen), with oligo-dT priming. After *AluI* and *RsaI* digestions and adaptor ligations, the constructs were PCR-amplified (GeneAmp Kit, Perkin Elmer, Foster City, CA). The amplified CD cDNA were digested with *EcoRI*, photobiotinylated (driver cDNA) and hybridized at 20-fold molar excess to an aliquot of non-biotinylated R cDNA. The hybridized non-specific sequences were eliminated by subsequent Streptavidin chromatography. After 3 cycles of selection, the amplified expressed sequence tag (EST) pool of the androgen-induced shutoff AS (R cDNA pool minus CD cDNA pool) sequences was digested with *EcoRI*, cloned into the BlueScript SK vector (Stratagene, La Jolla, CA) and transformed into E. coli (OneShot strain, Invitrogen).

Isolation of unique cDNAs from the differentially expressed sequence pool was performed as follows. Recombinants were collected randomly from the shutoff-positive AS pool of the Wang-Brown differential library and were plated. Using the labeled CD- and R-subtracted (CD cDNA pool minus R cDNA pool), PCR-amplified DNA master mixes as probes, double hybridizations revealed 11 and 14 clones that were present exclusively in the CD and R clone sets, respectively (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218). Multiple cross-hybridizations identified ten unique inserts.

To sequence the identified EST fragments, PCR sequencing reactions were performed using the dsDNA Sequencing System (Life Technologies, Gaithersburg, MD). The EST DNA sequences were tested for homology to known DNA sequences using the FASTA and BLAST (National Center for Biotechnology Information, Bethesda, MD) programs. Five inserts were found with no match in GenBank (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218). For further analysis, the mRNA with the highest induction in shutoff positive LNCaP-FGC cells (AS3, >5-6-fold of the 5.3 kb mRNA, and >3-4-fold of the 8 kb isoform) was selected.

To isolate the full length AS3 cDNA sequence, a 262 bp AS3 tag sequence was utilized to design nested primer pairs to amplify the full length cDNA sequence from a cDNA library. The cDNA libraries were generated by Human Genome Sciences (Rockville,

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MD), using polyA<sup>+</sup> mRNA preparations from androgen-treated or CDHuS-treated proliferation-arrested LNCaP-FGC cells. The Lambda ZAPII (UniZAP) phage was used as vector carrying *EcoRI* and *XhoI* cloning sites. The PCR reaction was designed to amplify the cloned unknown cDNA segments between the known tag sequence and the flanking vector sequences. Since the orientation of the tag sequence was not known, both ends of the insert were amplified in both directions. The vector primers used were commercially available sequencing primers: M13 Reverse and T3 primers at the *EcoRI* site, and M13-20 and T7 primers at the *XhoI* site.

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10 For the PCR reaction, the Expand High Fidelity kit was used and a 1 µl phage suspension as template (Boehringer-Mannheim). A 40 cycle amplification in a Perkin-Elmer 9600 thermocycler resulted in the production of a 1370 bp 5' fragment and a 3250 bp 3' fragment. These PCR products were purified using Qiagen columns, and sequenced by automatic sequencing using a primer walking strategy. The sequencing data showed that the open reading frame in the 5' end fragment did not have an authentic AUG codon.

15 To search for the missing 5' end of the transcript, the Prostate Specific Marathon Ready cDNA preparation from Clontech was used. Amplifications with the Clontech anchored primer and a set of AS3 specific primers resulted in a 419 bp fragment. The DNA was cloned and sequencing data showed that it carried the N-terminal 118 amino acids of the open reading frame. The nucleotide sequence reported herein has been submitted to  
20 GenBank under the accession number U95825 (see also, Geck *et al.*, (1999) *J. Steroid Biochem. Mol. Biol.* 68:41-50).

## EXAMPLE 2.

### CHARACTERIZATION OF THE AS3 cDNA SEQUENCE

25 In this example, the features of the AS3 mRNA and cDNA sequences are described.

Computer analysis of the sequenced 5253 bp AS3 cDNA identified a long open reading frame (Figure 1). The initiator methionine is at position 66, the stop codon was found at position 4239, and the region codes for a polypeptide of 1391 residues. The initiator is the first AUG codon downstream from the 5' end of the sequence, and appears in  
30 a strong Kozak-context (Kozak, (1991) *J. Biol. Chem.* 266:19867-19870; Kozak, (1991) *J. Cell Biol.* 115:887-903). The Northern blot size of the transcript is between 5.3 and 5.5 kb (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218) and since the sequence

reported here has 5253 nucleotides plus the poly-A tail, the 5' end of our sequence is at or within a few nucleotides of the 5' physical end, further suggesting that the initiator is authentic. The 5' non-coding region is high in GC nucleotides (63.3 %), but it has no recognizable secondary structure elements or other sequence features. The 3' noncoding region has several destabilizing AT-rich elements (underlined in Figure 1), typical of transcripts claimed to play a role in cell proliferation (Shaw *et al.*, (1986) *Cell* 46:659-667, Chen *et al.*, (1995) *Trends Biochem. Sci.* 20:465-470). The polyadenylation signal of the transcript is 25 bp upstream of the consensus GT-rich cleavage site (indicated in a larger font in Figure 1).

10

## EXAMPLE 3.

CHARACTERIZATION OF THE AS3 POLYPEPTIDE SEQUENCE

In this example, various structural and functional features of the AS3 polypeptide are described:

15 Computer analysis of the AS3 open reading frame was performed using the Translate program of the Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, Wisconsin.  $\beta$ -strand and  $\alpha$ -helix structures were calculated by the Chou-Fasman method using PepStructure and PepPlot programs. Motif and profile predictions were calculated using various programs of the Wisconsin Package, or by using remote servers offering sequence analyses of protein functional domains through the Internet. The following remote servers were used: PROWEB (<http://www.proweb.org>); BLOCKS (<http://www.blocks.fhcrc.org>); PRODOM (<http://www.toulouse.inra.fr/prodom/>); PRINTS (<http://www.biochem.ucl.ac.uk/cgi-bin/attwood/>) and the Protein Kinase Resource (<http://www.sdsc.edu/Kinases/>).

25 Employing the above programs, the expected molecular weight of the AS3 polypeptide is determined as 186 kD. In addition, it's noted that the N-terminal 400 amino acid domain is characterized by a unique arrangement of 31 aliphatic residues (21 of them are leucines). Every seventh position (with minor variations) is occupied by a leucine or similar hydrophobic residues and in the five subdomains shown in Figure 2, the pattern is uninterrupted. The arrangement is typical for coiled-coil structures where one side of the long  $\alpha$ -helices is hydrophobic and usually participates in protein-protein interactions (Lupas, (1996) *Trends Biochem. Sci.* 22:375-382; Beavil *et al.*, (1992) *Proc. Natl. Acad. Sci. USA*

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89:753-757). The leucine-zipper motif of DNA binding proteins is a specific subclass of this general pattern and the subdomain between positions 196 and 217 in the AS3 sequence is a perfect leucine-zipper.

The AS3 polypeptide sequence between positions 400 and 600 has elements of a conserved Mg-ATP binding domain of various nucleotide triphosphate binding proteins including protein kinases. In Figure 3, the AS3 sequence is shown in the conserved subdomain arrangements established by Hanks (Hanks *et al.*, (1991) *Methods Enzymol.* 200:38-62). The conserved  $\beta$ -strand,  $\alpha$ -helix structures and highly conserved critical residues are also indicated, together with the corresponding sequences of various protein kinases (Taylor *et al.*, (1992) *Annu. Rev. Cell Biol.* 8:429-462). Although the complete AS3 sequence did not appear to be related to any particular protein kinase or ATP binding protein, partial homology within the subdomains was maintained, and probably indicates that the domain is functional. Indeed, one feature of the AS3 polypeptide is the presence of several relatively well conserved kinase-related domains such as an Mg-nucleotide triphosphate binding pocket, as well as elements of the catalytic domain of several protein kinases (see Figure 3). Functional analysis of this domain using GST-fusion constructs indicates that the fusion construct polypeptide can form a complex resulting in the phosphorylation of at least two substrate proteins found in LNCaP-FGC cell extracts.

In addition to the above-mentioned motifs, the AS3 polypeptide also has a functional protein kinase domain located at amino acid residues 474-680 (encoded by nucleic acids 1420-2040; see SEQ ID NO: 3). To demonstrate that this region of the AS3 polypeptide has kinase activity, and is, e.g., capable of phosphorylating cellular substrates, the nucleic acid encoding this region was PCR amplified (using a corresponding upstream primer with a BamHI site and a corresponding downstream primer with a EcoRI site), digested, purified, and cloned into BamHI sites of the bacterial GST-fusion vector pGEX-T2 thereby fusing the AS3 domain to the C-terminus of GST (thus, the fusion protein is referred to as GST-AS3). The resultant construct, encoding a GST-AS3 fusion protein, was then transferred into bacteria (BL21 protease minus host) which were then induced to express the GST-AS3 fusion protein. Bacterial extracts containing the GST/AS3 fusion protein were incubated with MCF7-AR1 cell extracts in the presence of  $^{32}\text{P}$ -ATP and chromatographed through glutathione-Sepharose. Proteins which specifically bound to the GST-AS3 fusion protein were resolved by SDS-PAGE and autoradiography using standard conditions. Two polypeptides (40 and 120 kDa)

were specifically purified and phosphorylated by the AS3-GST fusion protein demonstrating the functional kinase activity of this protein kinase domain of AS3. There was no  $^{32}\text{P}$ -labelled band observed at 90 kDa (the mass of the AS3-GTS fusion protein) suggesting that the AS3 kinase domain does not auto-phosphorylate but phosphorylates other proteins through, e.g., a docking mechanism.

Furthermore, a putative nuclear localization sequence (NLS) (KKFTQVLEDDKIRK) resembling that of the androgen receptor and DNA polymerase- $\alpha$  was localized at position 547 (Zhou *et al.*, (1994) *J. Biol. Chem.* 269:13115-13123; Bouvier *et al.*, (1995) *Mol. Biol. Cell* 6:1697-1705). Further, the C-terminal region of the putative AS3 polypeptide contains several sequence elements that show similarities to DNA binding proteins. Motifs and ProfileScan searches in the Wisconsin Package indicated helix-loop-helix and Homeo-box signature sequences in the area, and a remote search on the BLOCKS server also identified DNA binding block elements in the C-terminal sequences. Still further, it is noted a serine-rich domain at position 1139, and a proline/glycine-rich domain at the 1284 position were also found. The C-terminal domain (about 200 amino acids) is highly charged and arranged in unique repeats of seven alternating acidic and basic domains.

A BLASTP search performed on the GenBank database resulted in a single high score similarity with the bimD gene of the eukaryotic organism *Aspergillus nidulans*, where 50% of the amino acid sequence was functionally similar in portions of the coiled-coil domain and the putative DNA binding domain at the C-terminus. The bimD protein has a basic leucine-zipper and a C-terminal charged (acidic) domain, similar to AS3, and appears to function as a DNA binding protein (Denison *et al.*, (1992) *Genetics*, 134:1085-1096). Both the AS3 and the bimD proteins also have nuclear localization consensus sequences.

Finally, to confirm that the AS3 polypeptide is capable of binding to DNA and moreover, to elucidate a DNA motif that binds to the DNA recognition sequence of AS3, an AS3-GST fusion polypeptide containing the DNA recognition sequence of AS3 was produced using a similar approach as used above to assay AS3 kinase activity.

Briefly, an expression vector was engineered to encode the DNA binding domain of AS3 fused to the N-terminus of GST (thus, the fusion protein is referred to as AS3-GST) which, when expressed in bacteria, allowed for the production of an AS3-GST fusion protein for DNA binding studies. The AS3-GST protein was then purified by affinity chromatography

as described above. Next, the resultant, purified AS3-GST protein was incubated with a mixture of oligonucleotides comprising constant 5' (25 bp) and 3' (25 bp) ends and randomly generated middle segments (18 bp) and chromatographed through a glutathione-Sepharose the column. To avoid non-specific binding, double stranded poly(dI-dC) (average length 1800 bp) was added to the mixture. The AS3-GST fusion protein, together with bound oligonucleotides, was eluted with glutathione and the bound oligonucleotides were amplified by PCR, purified, and subjected to a second round of column chromatography using AS3-GST before being PCR amplified and cloned into a TA vector.

Thirty clones were sequenced to assess whether a specific nucleotide sequence was recognized by the AS3-GST fusion product. The sequences were compared and analyzed by the PILE-UP program of the GCG package. The co-alignment of the sequences revealed a pattern of the following putative consensus sequence:

5' C, T, A, [T/A], [T/A], A, G, [C/G], C, C, C, [C/G], G, C, [C/G], C, A, [A/T], 3' (SEQ ID NO: 5)

Interestingly, sequence similarities were found between the putative AS3 DNA recognition sequence and the NF-kappaB and the Mbp1 recognition motifs. In addition, similarities within the p27<sup>kit1</sup> promoter at positions -270, -383, -427, and -586 were also found indicating that important gene regulatory elements exist in the cell in which the AS3 protein may interact with (Zhang *et al.*, (1997) *Biochem. Biophys. Acta.*, 1353:307-317). Interestingly, the transcription factor NF-kappaB, which is involved in the control of apoptosis, and Mbp1 which is involved in the G1-S cell cycle transition in yeast, also interact with similar DNA motifs (Waddick *et al.*, (1999) *Biochem. Pharmacol.*, 57:9-17; Koch *et al.*, (1993) *Science*, 261:1551-1557).

Accordingly, it was concluded that AS3 is capable of binding to DNA in a sequence specific manner and therefore, may function as a transcriptional regulator of genes involved in cell growth control.

## EXAMPLE 4.

THE AS3 GENOMIC LOCUS AND USE OF AS3 RELATED MOLECULES AS  
MARKERS FOR DISEASE

5 In this example, aspects of the AS3 genomic locus and the use of AS3 related molecules as markers for disease are described.

A computer homology search in GenBank was performed and identified the AS3 genomic region as residing on chromosome 13q12-q13. This area is represented by cosmid 267p19, and on P1 artificial chromosomes PAC26H23 and PAC49J10. Consensus splicing  
10 donor and acceptor sites were identified and the entire exon-intron structure of the AS3 gene was resolved comparing the cDNA sequence and the genomic sequence using the BLAST program (see Figure 4) (Shapiro *et al.*, (1987) *Nucl. Acids Res.* 15:7155-7174). The actual cosmid and cDNA positions are listed in Figure 4, and the arrangement of exons is depicted in Figure 5. The area covers nearly 200,000 bp and the average size of the exons is 100-150  
15 bp.

Interestingly, the AS3 genomic area is centromeric to the RB1 locus, and telomeric to BRCA2. The AS3 gene is transcribed in the same direction as BRCA2, and the coding strand is downstream from the breast cancer gene (Couch *et al.*, (1997) *Genomics* 36:86-99). On the opposite strand upstream of AS3, three regions were assigned to cDNAs  
20 of unknown functions. An expressed sequence (CG008) has been assigned to this area, and represents a portion of the AS3 transcript (Couch *et al.*, (1997) *Genomics* 36:86-99). The N terminal 354 amino acids of the open reading frame are missing in the CG008 sequence in GenBank. The CG008 open reading frame terminates at amino acid 738 of the AS3 sequence. The sequencing data reported herein and the published genomic sequence are  
25 identical, confirming the correct sequence of AS3. The extra C at nucleotide position 1,109 in the CG008 sequence suggests a possible sequencing error that results in a frame shift and a stop codon at position 1,152 of the CG008 sequence.

With the above information in hand, it was observed that several epidemiological studies support a link between breast and prostate cancers implying shared  
30 genetic suppressor elements in both disease states (Thiessen, (1974) *Cancer* 34:1102-1107; Tulinius *et al.*, (1992) *Br. Med. J.* 305:855-857). For example, studies of breast cancer families with high loss of heterogeneity (LOH) in the BRCA2 area showed that high



prostate cancer incidence also occurred in 4 out of 5 families examined (Gudmundsson *et al.*, (1995) *Cancer Res.* 55:4830-4832). Further, in the majority of the male relatives with prostate cancer in these families (86%), allelic losses in the BRCA2 area were also detected with some of these mutations occurring in the region immediately downstream of the

5 BRCA2 gene (Gudmundsson *et al.*, (1995) *Cancer Res.* 55:4830-4832; Van den Berg *et al.*, (1996) *Br. J. Cancer* 74:1615-1619; Cleton-Jansen *et al.*, (1995) *Br. J. Cancer* 72:1241-1244).

Moreover, it was noted that putative suppressors in the immediate vicinity of BRCA2 are not only limited to sex hormone-related cancers. For example, recent studies on

10 chronic lymphoid leukemia detected a 1Mb allelic loss in this region, with no mutations in the BRCA2 gene, pointing to a cryptic suppressor next to this gene (Garcia-Marco *et al.*, (1996) *Blood* 88:1568-1575; Caldas *et al.*, (1997) *Proc. Am. Assoc. Cancer Res.* 38:191). As indicated herein, the coding sequence of AS3 lies within this area. Thus, AS3 (or AS3-related molecules) may be associated with a number of diseases and conditions at several

15 different levels involving, e.g., genomic alterations (including deletions and/or mutations at the chromosomal level), altered transcription or transcript production, and/or altered protein or protein expression levels. Thus, "AS3 related molecules" include, nucleic acid fragments or probes derived from the genomic locus (or an AS3 cDNA), AS3 or AS3 related proteins or protein fragments.

20 In addition to foregoing, the ability to determine AS3 protein expression levels, altered AS3 proteins, and/or AS3 protein expression patterns in, for example, different tissue samples (such a biopsy sample from, e.g., the prostate) may be desired. To this end, several antibodies have been developed that specifically bind the AS3 polypeptide. Briefly, computer aided sequence analysis of the AS3 protein was performed in order to identify several antigenic areas

25 of the AS3 protein that were suitable for use as an immunogen. Accordingly, oligopeptides corresponding to amino acid residues 711-727 and 1,369-1387 of SEQ ID NO: 4 were synthesized and used to raise antibodies in, respectively, rabbits and chickens (egg yolk immunoglobulin Y). The resultant antibodies were both tested against cellular extracts derived from either LNCaP-FGC or MCF7-AR1 cells induced with androgen to express AS3 protein

30 and both antibody preparations recognized a protein band of the expected size for AS3 protein. Thus, these antibodies can be used for performing, for example, immunocytochemistry on

various cell samples (e.g., biopsies of the prostate) for various prognostic and diagnostic determinations.

Accordingly, it was concluded that the AS3-related molecules and AS3-specific antibodies disclosed herein are useful prognostic/diagnostic probes for evaluating diseases (e.g., the aforementioned cancers) that are associated with, or map to, the AS3 locus or are associated with altered AS3 protein expression.

#### EXAMPLE 5.

##### USE OF AS3 MOLECULES IN THE TREATMENT OF PROSTATE CANCER

10 In this example, the use of AS3 as a marker in guiding the appropriate administration of hormone therapy for prostate cancer is discussed.

It was determined that expression analysis of the AS3 transcript demonstrated proliferation arrest-specific induction patterns, starting soon (4-6 h) after androgen exposure (Geck *et al.*, (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218). AS3 levels peaked at 18-  
15 20 h, about 4 h before the commitment for proliferative shutoff was detected, suggesting that this gene is a candidate for a shutoff mediator. Furthermore, expression of the AS3 transcript positively correlated with proliferation arrest as this gene was expressed only in shutoff-positive cell lines and variants. In addition, LNCaP-FGC cells proliferated maximally in CDHuS supplemented with 30 pM R1881 and under these conditions AS3 was  
20 not expressed. When AS3 was strongly induced in the presence of hormone (i.e., 0.3-30 nM of R1881), the cells were inhibited from proliferating. An additional observation that indicates the AS3 gene codes for an inhibitor of the proliferation of prostate cells is the increase of AS3 mRNA levels in the rat prostate when proliferation was arrested by prolonged androgen administration. In addition, comparable effects in MCF7-AR1 cells  
25 have been observed. Finally, significant homology between the fungal gene bimD and AS3 was found. Importantly, it was noted that overexpression of bimD in *Aspergillus nidulans* results in a cell cycle arrest in G1/S phase and observe a similar cell cycle arrest and concordant peak expression of AS3 in mammalian cells induced to undergo androgen-induced proliferative shutoff (Denison *et al.*, (1992) *Genetics*, 134:1085-1096; Geck *et al.*,  
30 (1997) *J. Steroid Biochem. Mol. Biol.* 63: 211-218).

Based on these observations, it was concluded that AS3 expression is a useful marker of responsiveness of prostate cancer cells to the inhibitory effect of androgens. In

order to make the tumor regress, avoid reoccurrence, and maintain an acceptable quality of life, patients presenting this response may be treated with alternate cycles of antiandrogens and androgens. Androgens have a biphasic effect on the normal prostate: an initial phase of increased proliferation (Step 1) followed by a phase of inhibition (Step 2).

- 5 Prior to the invention, prostate cancer has been treated by hormone ablation (castration, antiandrogens) to take advantage of Step I. Usually after a significant regression, the remaining tumor cells become resistant and the tumor and/or metastases relapse. The invention avoids this problem by allowing the clinician to subject patients to an intermittent therapy. This is based on a classification of patients who have an increased
- 10 chance to be responsive to the intermittent hormone therapy. Patients not showing AS3 positive cancer cells would then be subjected to alternative therapies (chemotherapy, radiation, etc.). In contrast, the protocol for AS3 positive patients would be to first administer antiandrogens, to block the proliferative effect and return PSA levels to normal, and then to treat the patient with physiological but high doses of androgen to elicit Step 2.
- 15 The invention allows the clinician to confidently assess the initial hormone dependence of the tumor (i.e., whether tumor cells express AS3 in response to hormone) and determine when to re-expose the patient to hormone. Importantly, the use of intermittent hormone therapy allows the clinician to determine exactly when cells are no longer responding to hormone such that hormone treatment can be withdrawn before the cells become refractory
- 20 to hormone treatment and untreatable. If the patient develops metastases capable of being biopsied, assaying AS3 levels as described herein would allow a renewed attempt to reduce the proliferation of those cancer cell by increasing the androgen concentration of the treatment. This important aspect of the invention should allow the clinician to lower the rate at which clones of cells develop hormone resistance and become capable of multiplying and
- 25 metastasizing without androgen stimulation. Moreover, this new treatment regime affords a better quality of life for the patient because, unlike with constant hormone treatment, sexual drive and potency are recovered.

## EXAMPLE 6

ANTISENSE ASSAY FOR DEMONSTRATING THAT AS3 IS AN ANDROGEN-INDUCED SUPPRESSOR OF CELL PROLIFERATION

In this example, an inducible antisense assay is provided for demonstrating that AS3  
5 is an androgen-inducible suppressor of cell proliferation.

To demonstrate androgen-inducible AS3 suppressor activity in a proliferating cell (i.e.,  
proliferative shut off activity), clonal cell lines were derived in which the presence or absence  
of AS3 could be controlled using an inducible AS3 antisense gene. The prediction was that  
cells incubated in the presence of androgen would have reduced levels of proliferation as  
10 compared to the same cells in the absence of androgen or cells in the presence of androgen but  
in which levels of AS3 had been experimentally ablated. Thus, cells incubated in the presence  
of androgen, but also induced to express antisense AS3 gene transcript which removes cellular  
levels of the AS3, would be predicted to grow like wild type cells because, even though the  
cells were being exposed to androgen, there would be insufficient AS3 to mediate the  
15 androgen signal reducing cell growth. This is precisely what was observed (see, e.g., Table 1),  
thereby demonstrating that AS3 is an androgen-induced suppressor of cell proliferation.

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→ In order to develop the foregoing novel cell lines in which to demonstrate that AS3  
mediates the androgen-induced shutoff effect, an inducible transgene encoding an AS3  
antisense transcript (or empty vector as a negative control) was genetically engineered into a  
20 retroviral vector backbone for efficient, stable, integration into cells. In particular, the AS3  
antisense gene was cloned into the Clontech pRevTRE retroviral vector under the control of  
a tetracycline sensitive promoter. The promoter has seven repeats of the bacterial *tetO*  
operator sequence upstream of the minimal CMV promoter which can be bound by the  
tetracycline transactivator (tTA). The tTA is a fusion protein between the bacterial  
25 tetracycline repressor and the V16 herpes virus transactivator. The tetracycline  
transactivator is sensitive for tetracycline such that, in the presence of tetracycline, the  
transactivator cannot bind the tetracycline promoter so the transgene is "off" and conversely,  
in the absence of tetracycline, the gene is "on" (i.e., the Tet-Off system; see, e.g., Clontech  
pRevTRE manual for further details).

30 To stably introduce the foregoing inducible vectors into cells, the vectors were first  
transfected into a packaging cell line (PT67) under drug selection (hygromycin). After 2-3

weeks of selection, the surviving packaging cells were used as a source of supernatants containing infectious virions containing the inducible transgene.

The supernatants containing the highest amounts of virions were then used to infect the STFX1 cell line, a derivative of the MCF7-AR1 that has a tetracycline transactivator gene stably integrated. During the cloning and selection procedures, cells were maintained in 1 µg/mL of tetracycline to suppress the expression of the inducible transgenes.

In order to characterize the cell lines expressing AS3 antisense and the "control" cell lines having vector without insert, cells were incubated in the presence (i.e., the transgene is off) and absence of tetracycline (i.e., the transgene is on) and, after 36 hours, assayed for transcript expression using RT-PCR under standard conditions,

To determine the affect of androgen induced AS3 suppression of cellular growth, cells lines showing high levels of inducible AS3 antisense expression were chosen for further study. Cells were seeded onto coverslips in standard growth medium and allowed to grow for 5 days in the presence of 10 µg/ml tetracycline (hereafter "tet"). Then, the medium was changed to i) 10 µg/ml tet, or ii) no tet for 36 hours as indicated in Table 1. Then, vehicle or 10 nM of the androgen R1881 was added. Finally, 24 hours later, the cells were treated with 10 µg/ml of bromodeoxyuridine (BrdU) for several hours to measure the percent of cells actively proliferating as a function of BrdU incorporation during DNA replication. The cells were then fixed, Hoechst stained, and the percent number of BrdU-labeled cells was determined using immunocytochemistry (using standard BrdU labeling reagents and protocols from Boheringer).

As clearly indicated in Table 1, administration of the androgen R1881 results in a significant decrease in the number of proliferating cells in S phase (compare to sample 1 where cells are not treated with androgen) when cellular levels of AS3 are unaffected because the transgene vector is empty and uninduced (sample 2), empty and induced (sample 4), or encoding AS3 but uninduced (sample 6). In stark contrast, when AS3 levels are reduced by antisense expression in the absence of tetracycline, the androgen R1881 no longer induces a proliferative shutoff (compare sample 8 with sample 6).

**Table 1. Analysis of the effect of tetracycline and androgen on the percent BrdU-labeled nuclei of STFX1 cells containing vector only or AS3 antisense constructs**

Sample	Treatment	% Proliferating Cells (Mean $\pm$ S.E.)
1	vector, +tet, -A	35.0 $\pm$ 1.7
2	vector, +tet, +A	9.4 $\pm$ 1.7
3	vector, -tet, -A	36.6 $\pm$ 2.7
4	vector, -tet, +A	10.7 $\pm$ 0.9
5	Antisense, +tet, -A	34.7 $\pm$ 1.0
6	Antisense, +tet, +A	9.7 $\pm$ 0.6
7	Antisense, -tet, -A	32.2 $\pm$ 1.5
8	Antisense, -tet, +A	29.1 $\pm$ 1.6

Vector denotes STFX1 cells expressing empty vector under tetracycline control. Tetracycline is abbreviated tet; A denotes 10 nM R1881. Antisense denotes STFX1 cells expressing AS3-antisense under tetracycline control. Tetracycline represses the expression of the vector and the AS3 antisense. Each data point represents the mean of 8-9 low power fields containing about 150-200 cells/field. Data were analyzed using the Mann-Whitney test; significance was measured at  $p < 0.001$ .

This experiment further shows that the expression of the empty vector neither interferes with the androgen-mediated shutoff, nor results in toxic effects.

Accordingly, it is concluded that androgen induced suppression of cell proliferation (or proliferative shutoff) is modulated by AS3 because expression of an AS3 antisense that reduces cellular levels of AS3 results in the blocking of androgen-induced AS3-mediated suppression of cell growth.

#### Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

What is claimed: